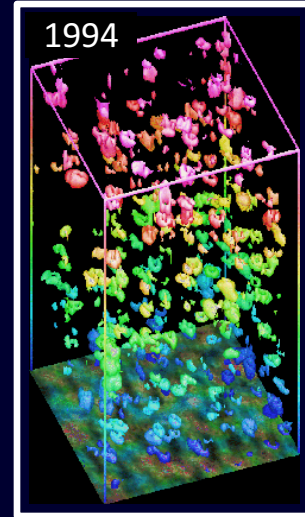
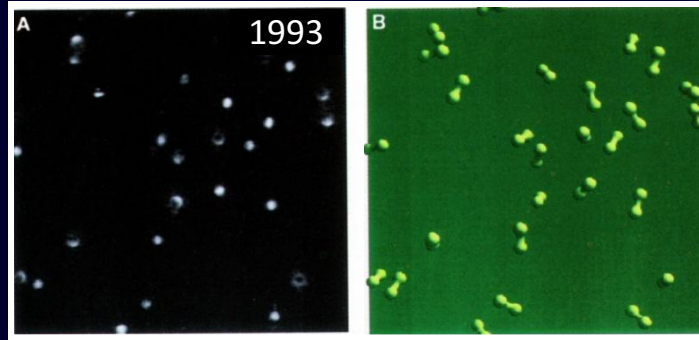
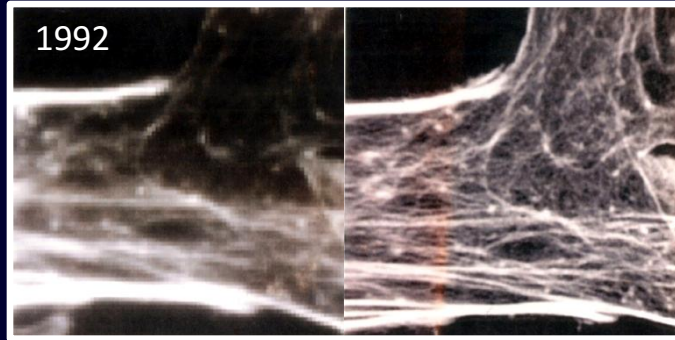


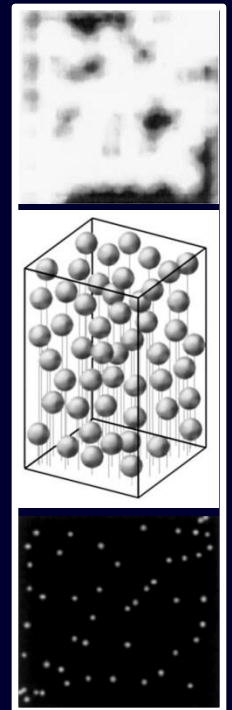
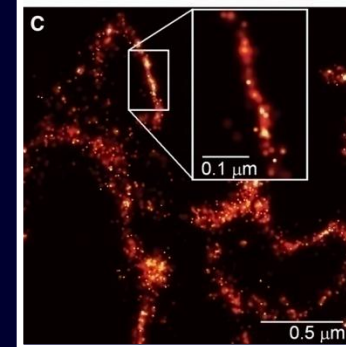
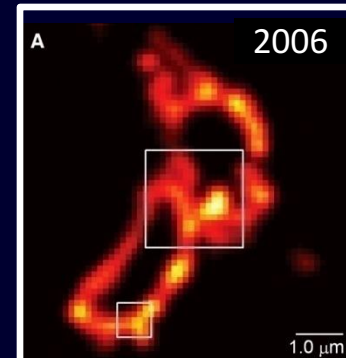
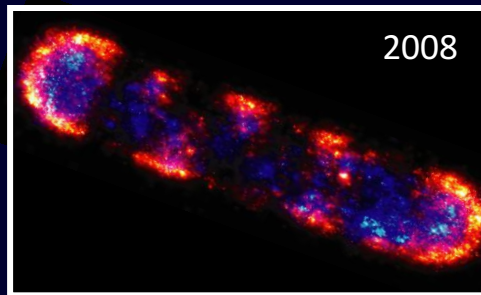
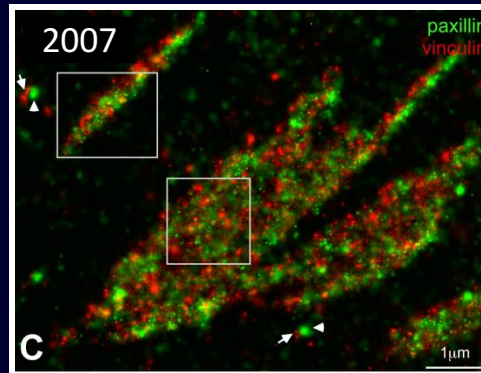
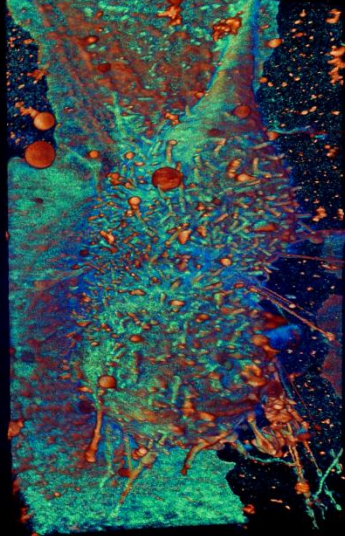
Single Molecules, Cells, and Super-Resolution Optics

Eric Betzig
Janelia Research Campus, HHMI



2014

LLCPK1 Cell Intracellular Membranes Plasma Membrane Histone H2B



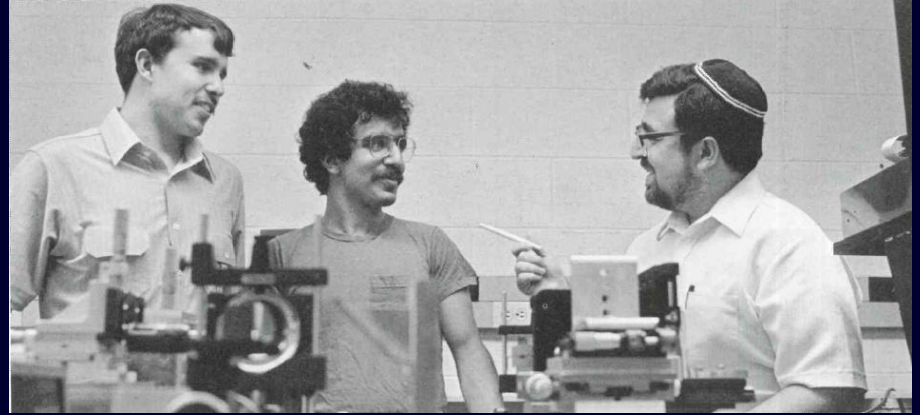
1995

Cornell and the Beginnings of Near-Field Optical Microscopy

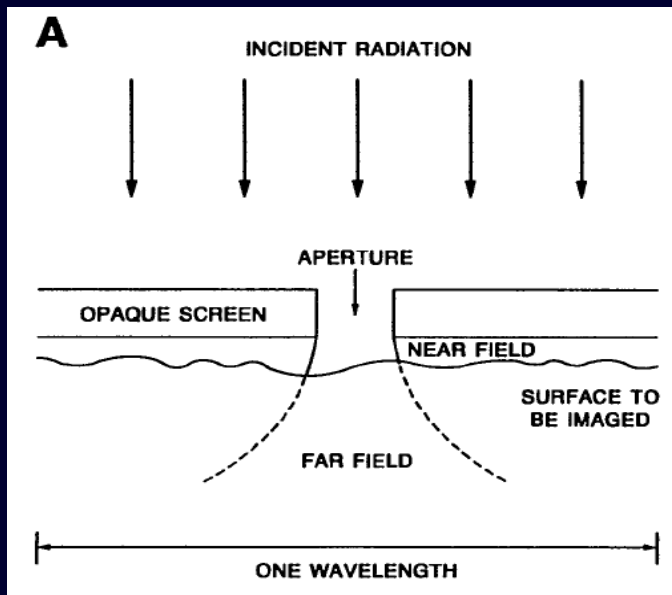
Mike Isaacson and his STEM



Me, Alec Harootunian, and Aaron Lewis, 1983



concept

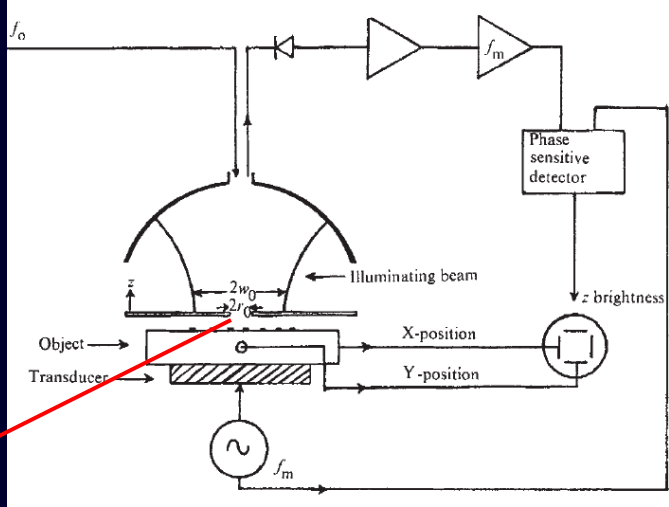
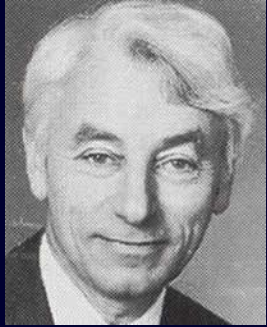


A. Lewis, *et al.*,
Ultramicroscopy 13,
227 (1984)

The Long History of Breaking Abbe's Law: Near-Field

near-field microwave ($\lambda = 3$ cm) microscopy

Sir Eric Ash



sub-wavelength aperture

object



image



Resolution of $1/60$ of the wavelength!

XXXVIII. A Suggested Method for extending Microscopic Resolution into the Ultra-Microscopic Region. By E. H. SYNGE*.



Edward "Hutchie" Synge, *Phil. Mag.* 6, 356 (1928)

- J.A. O'Keefe (1956)
- A.V. Baez (acoustics, 1956)
- C.W. McCutchen (1967)
- U. Ch. Fischer (lithography, 1981)
- D.W. Pohl (1984)
- G.A. Massey (1984)
- J. Wessel (1985)

The Long History of Breaking Abbe's Law: Far-Field

Structured Light

Optical Systems with Resolving Powers Exceeding the Classical Limit*

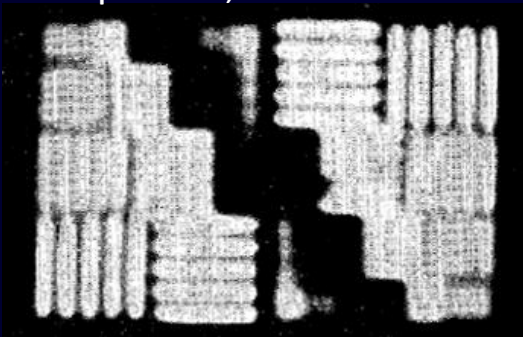
W. Lukosz†

Institut A für Physik, Technische Hochschule, 33 Braunschweig, Germany

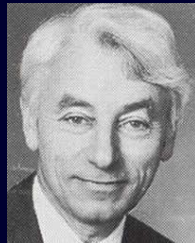
(Received 27 April 1966)

W. Lukosz, *JOSA* 56, 1463 (1966)

test pattern, conventional

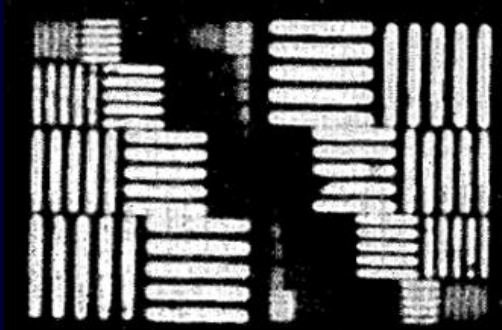


Sir Eric Ash



intentional overexposure

test pattern, super-resolved

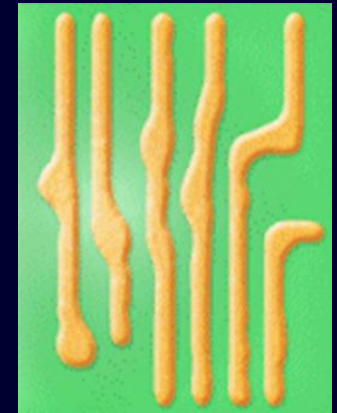
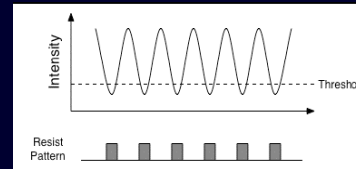
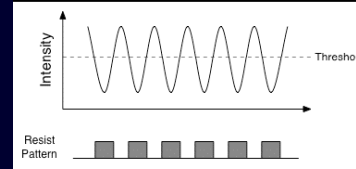


Resolution 3×
beyond Abbe's
Limit!

A. Bachl, W. Lukosz, *JOSA* 57, 163 (1967)

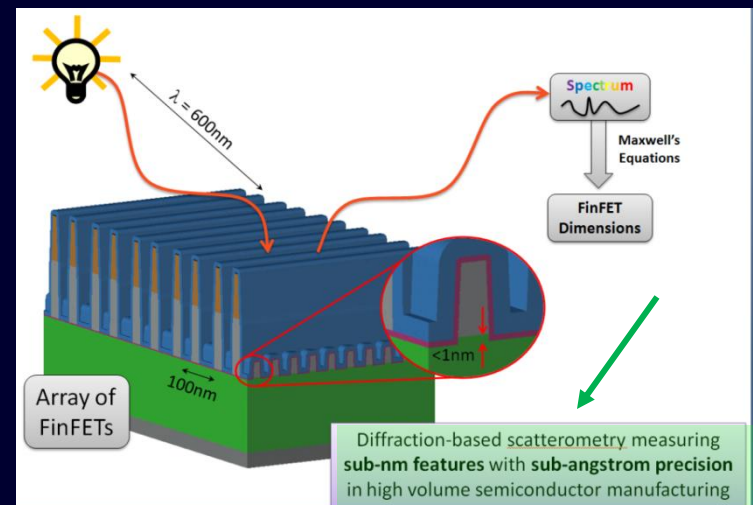
Nonlinear Interaction with Sample

integrated circuit linewidth control



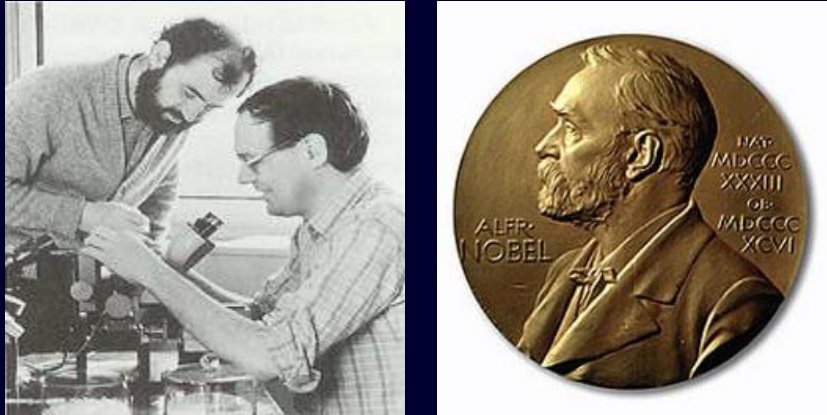
λ

A Priori Information: wafer inspection

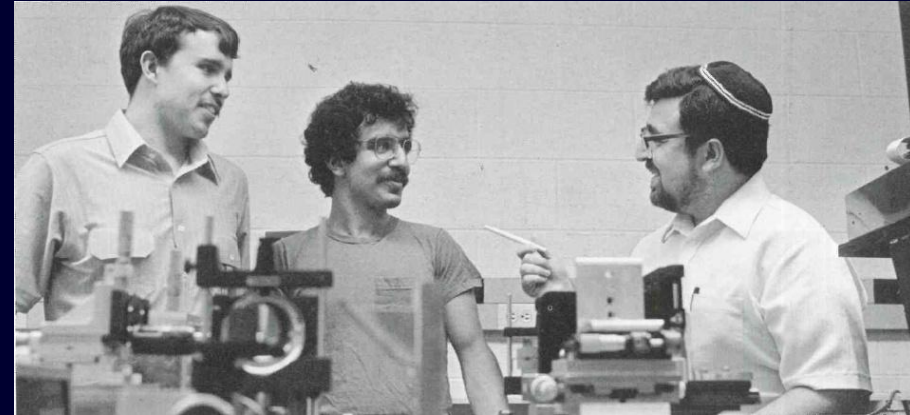


Making Near-field *Optical* Microscopy Work

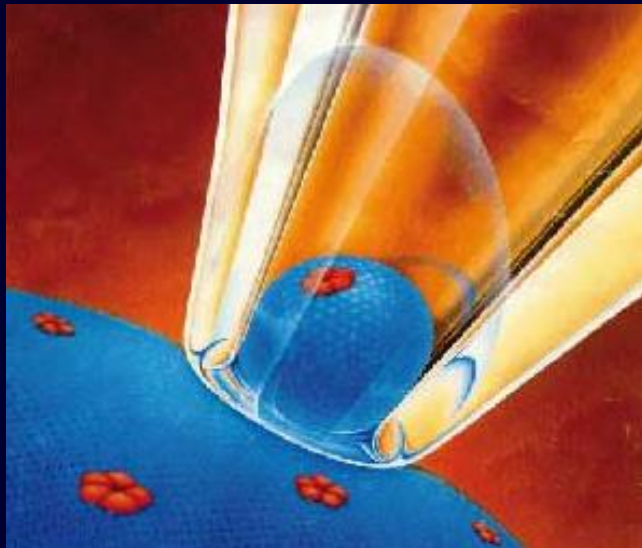
Edwin Neher and Bert Sakmann, Nobel 1991



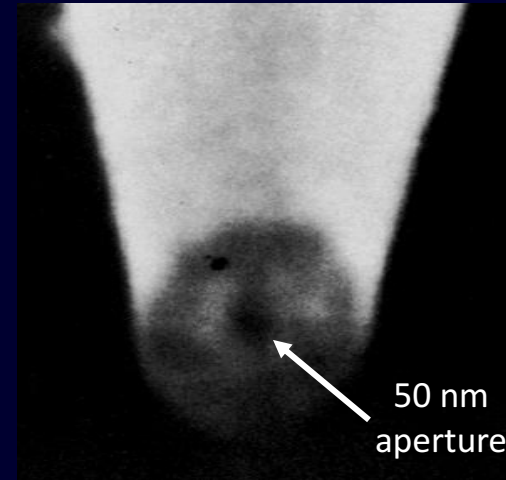
Me, Alec Harootunian, and Aaron Lewis, 1983



patch clamp: single ion channel recording



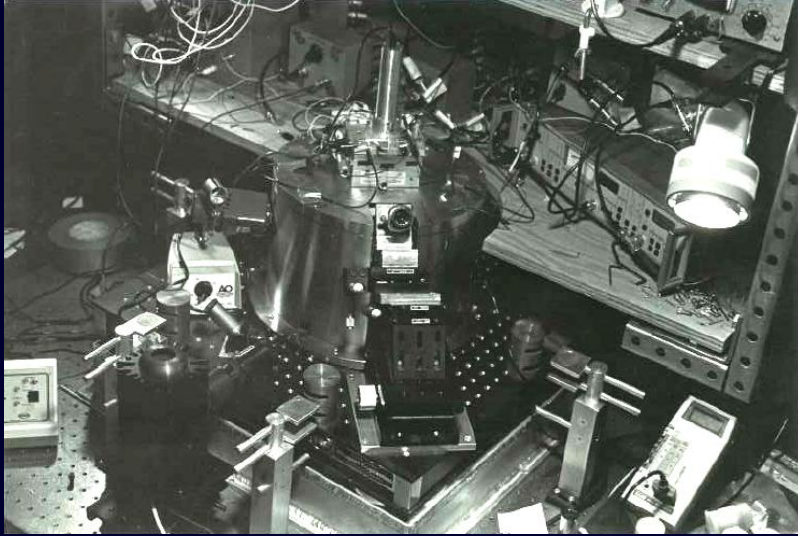
end of aluminum coated pipette



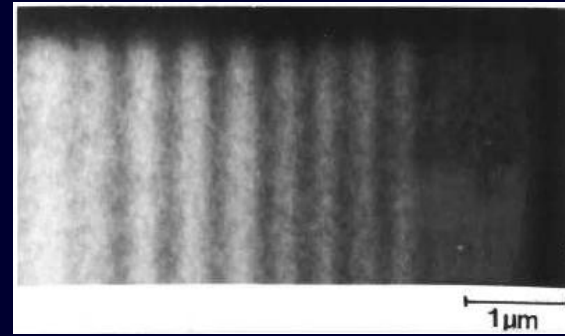
E. Betzig, *et al.*, *Biophys. J.* **49**, 269 (1986)

Making Near-field *Optical* Microscopy Work

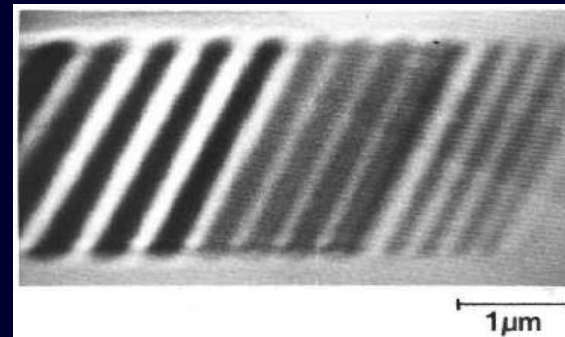
my near-field scanning optical microscope (NSOM)



microscope control room



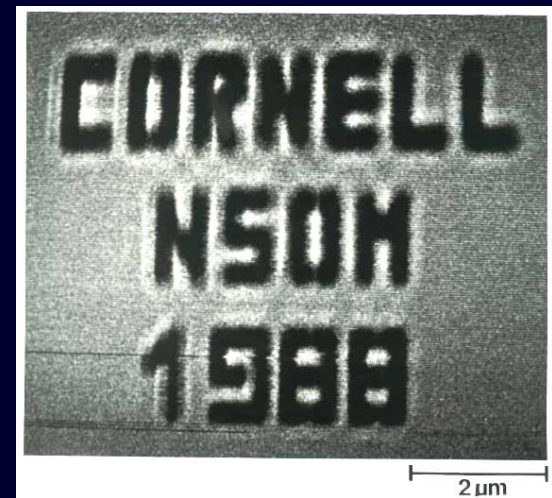
diffraction
limited



NSOM

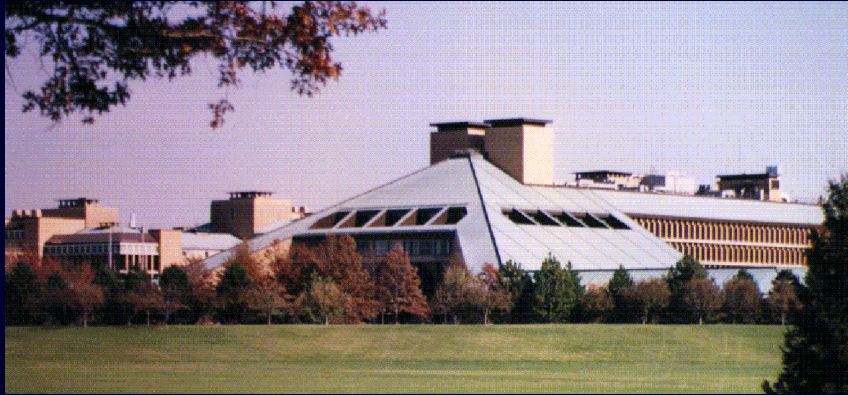


NSOM

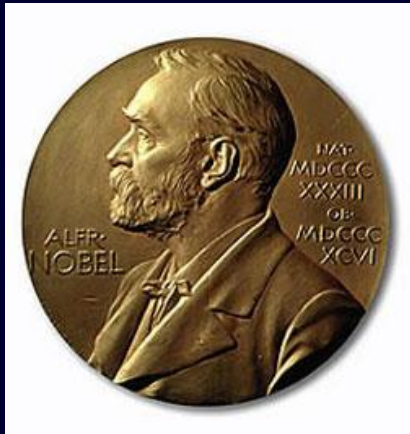
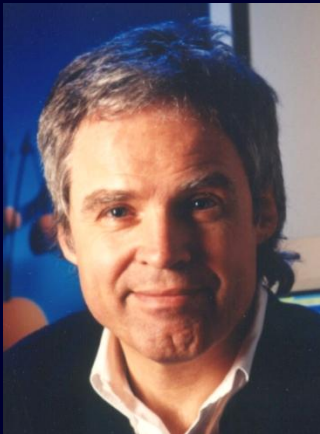


Initial Struggles at Bell Labs

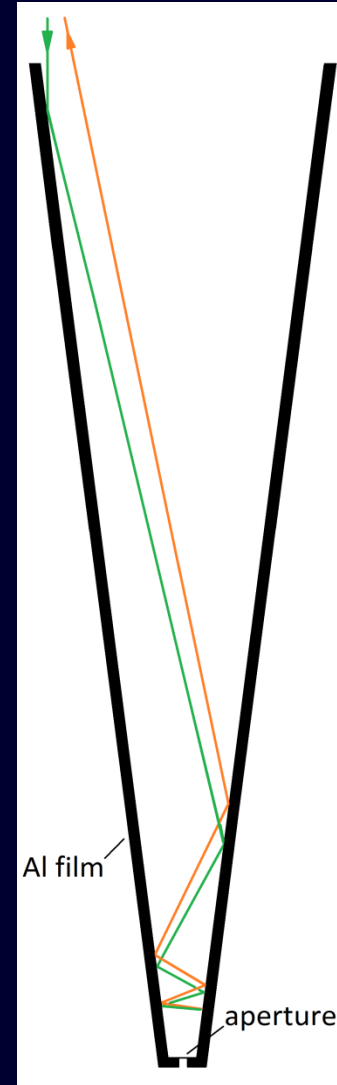
AT&T Bell Labs, Murray Hill, NJ



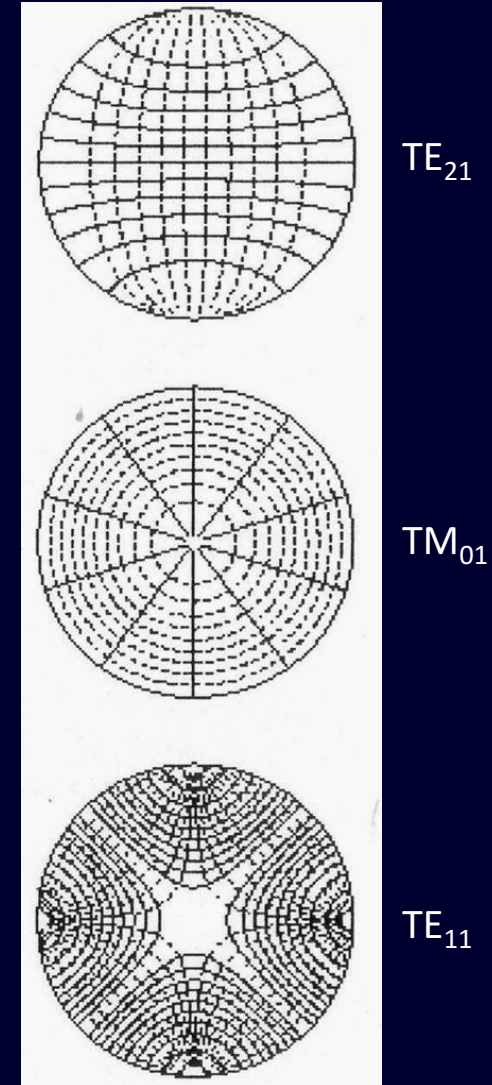
Horst Störmer, 1998 Nobel in Physics



retroreflection
in pipette

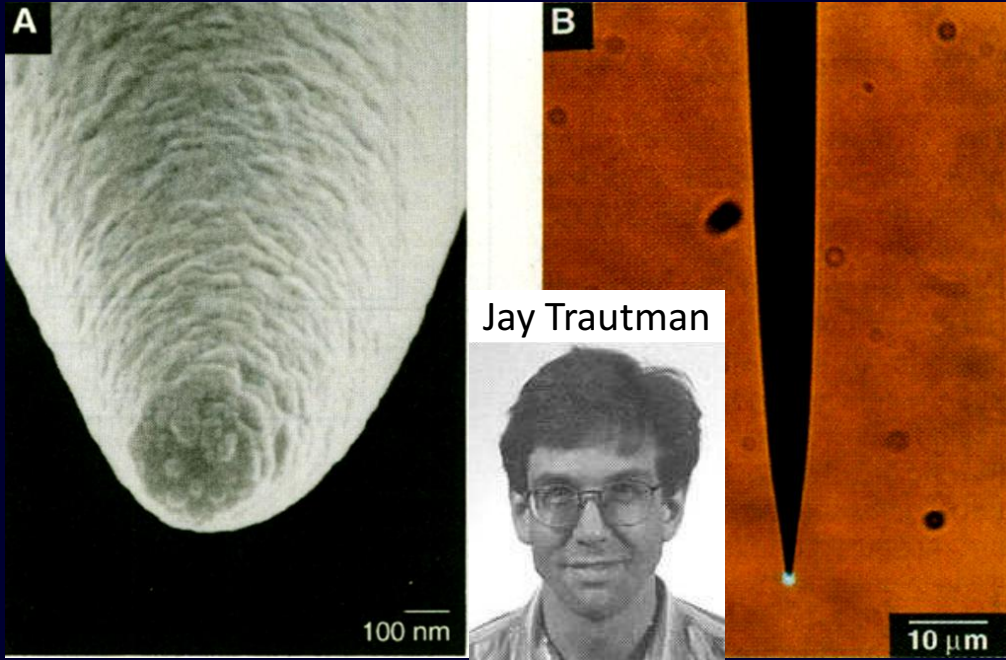


lowest order waveguide
modes at tip

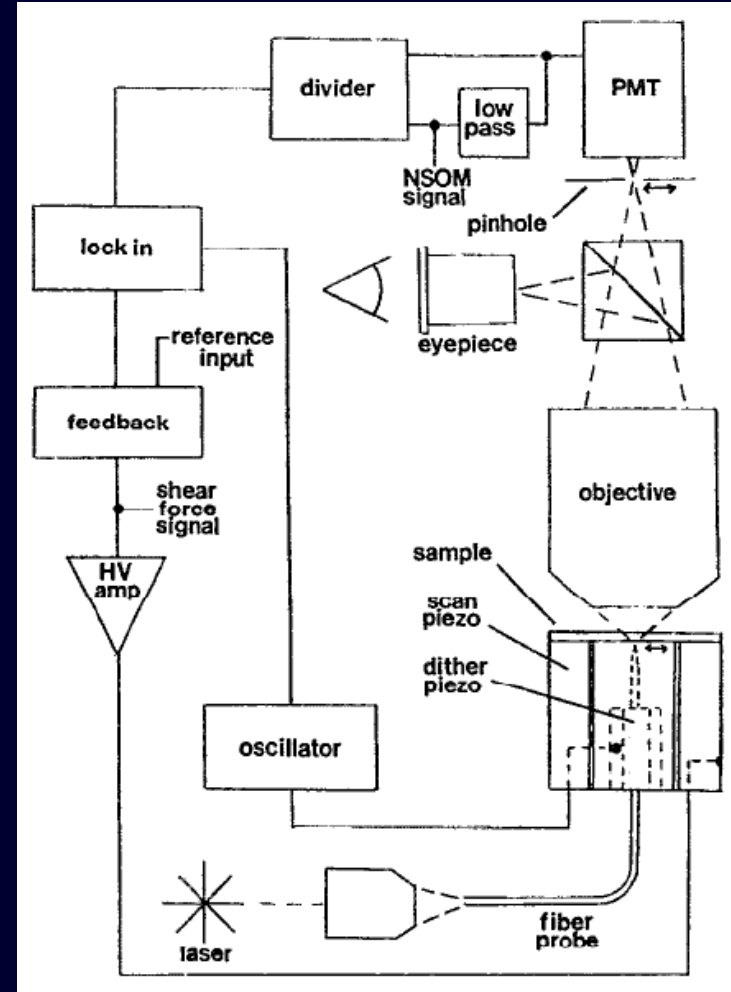


Making NSOM Routine

adiabatically tapered optical fiber probe



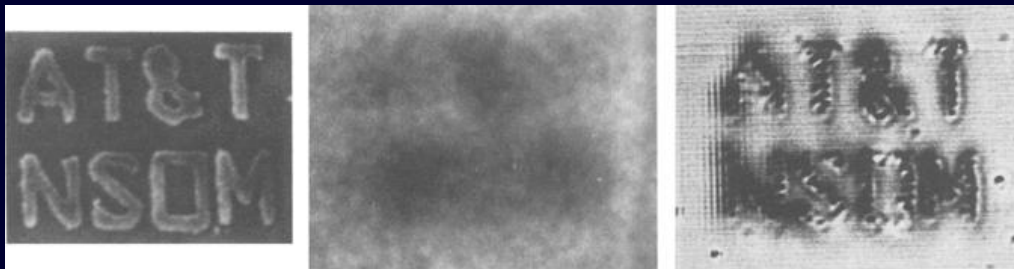
shear force distance regulation



SEM

widefield

NSOM



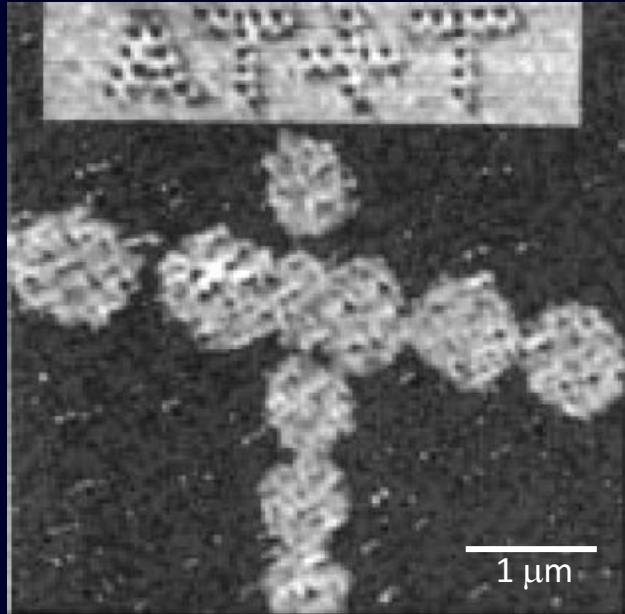
1 μm

E. Betzig, J.K. Trautman, *et al.*, *Science* **251**, 1468 (1991)

E. Betzig, *et al.*, *Appl. Phys. Lett.* **60**, 2484 (1992)

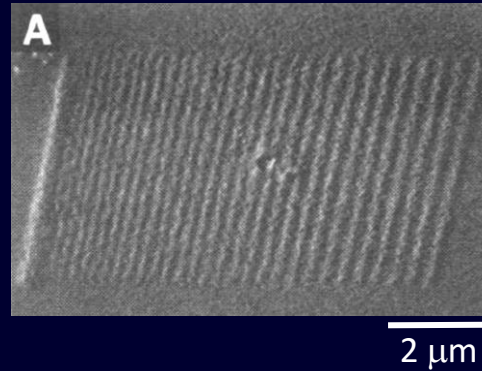
The Golden Age of NSOM

high density data storage

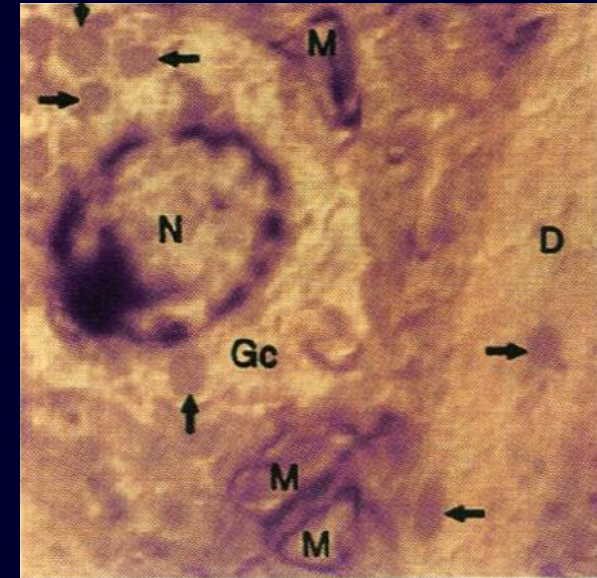


E. Betzig, *et al.*, *Appl. Phys. Lett.* **61**, 142 (1992)

photolithography

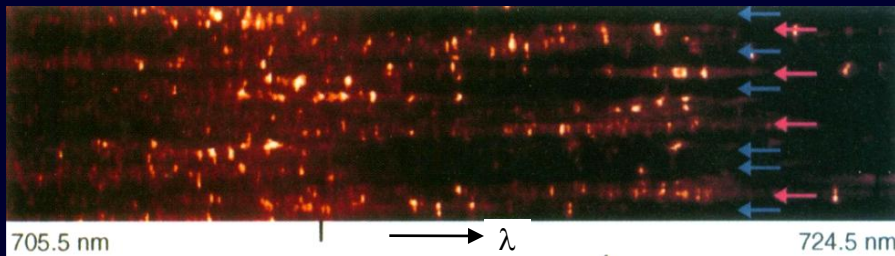


histological section,
monkey hippocampus



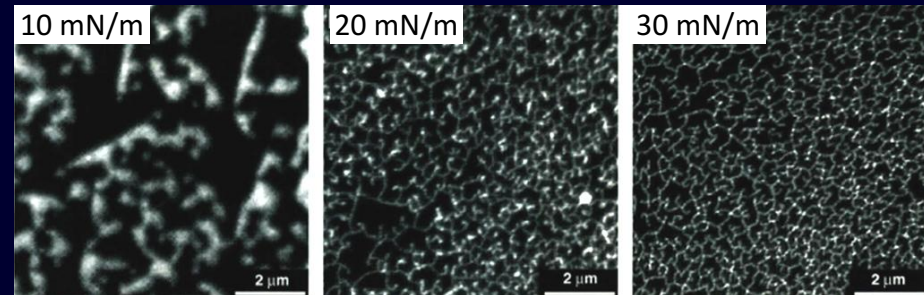
E. Betzig, J.K. Trautman, *Science* **257**, 189 (1992)

nanoscale spectroscopic imaging



H.F. Hess, *et al.*, *Science* **61**, 142 (1994)

fluorescence: phase change in phospholipid monolayers



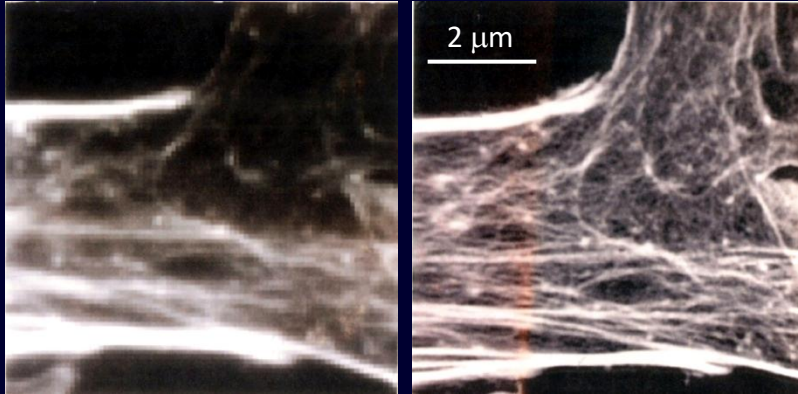
J. Hwang, *et al.*, *Science* **270**, 610 (1995)

Single Molecule Detection (SMD)

fluorescence: actin, mouse fibroblast cell

widefield

NSOM



E. Betzig, *et al.*, *Bioimaging* 1, 129 (1993)

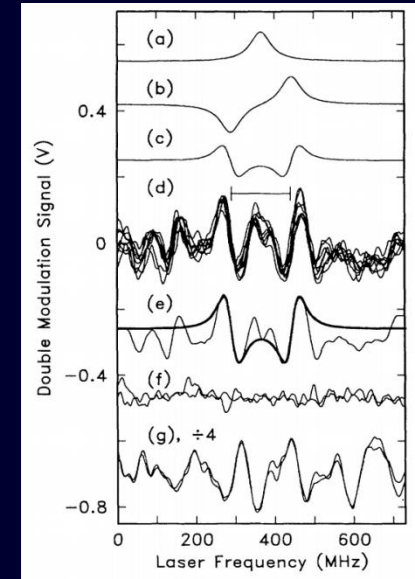
Nobel, 2014



W.E. Moerner

W.E. Moerner, L. Kador,
Phys. Rev. Lett. **62**, 2535
(1989)

single molecule absorption spectra, 1.6°K

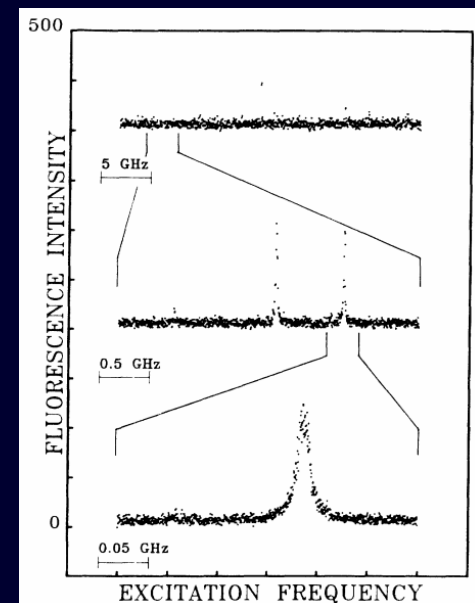


SM fluorescence excitation spectrum, 1.8°K

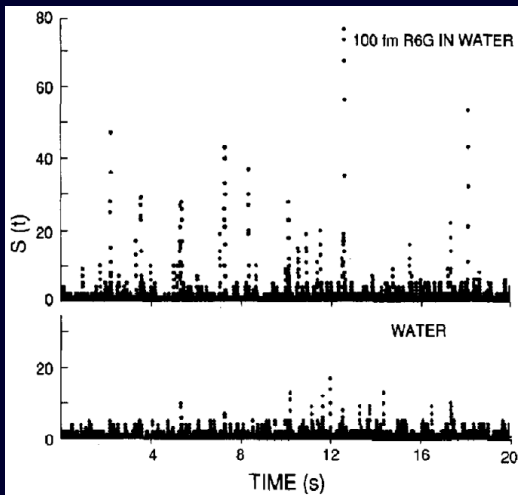


Michel Orrit

M. Orrit, J. Bernard,
Phys. Rev. Lett. **65**,
2716 (1990)



SM fluorescence bursts at room temp



Time gated:

E.B. Shera, *et al.*, *Chem. Phys. Lett* **174**, 553 (1990)

FCS:

R. Rigler, J. Widengren,
Bioscience **3**, 180
(1990)

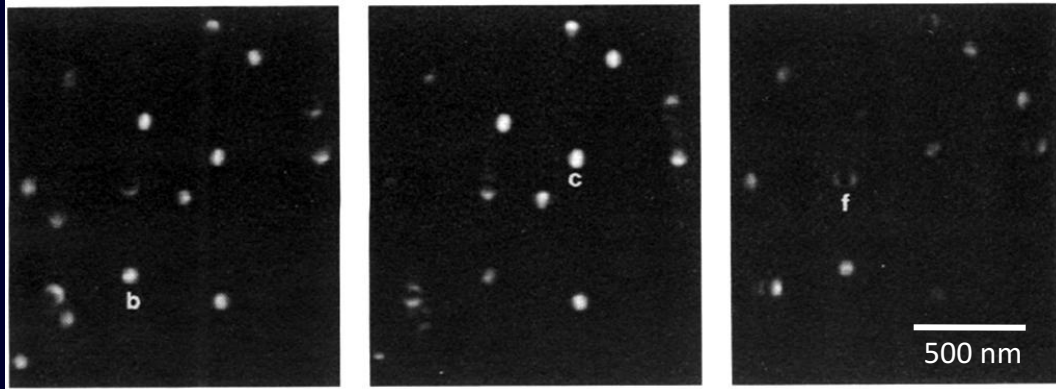
NSOM and the Birth of Single Molecule Microscopy



Rob Chichester

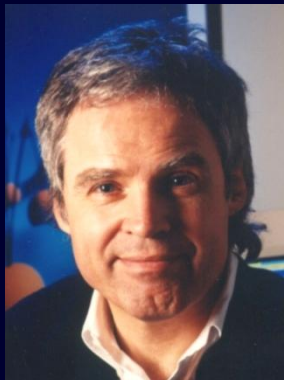
single molecule fluorescence anisotropy

random

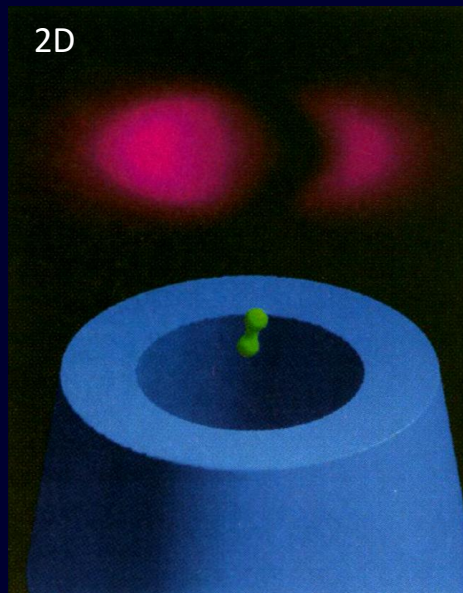
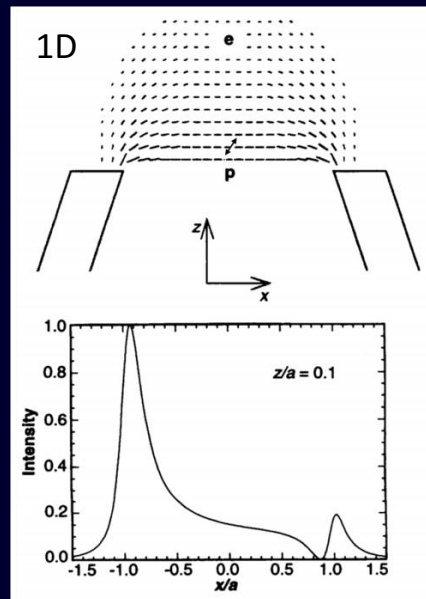


dil-C₁₈-(3)
molecules on
PMMA

E. Betzig, R.J. Chichester,
Science **262**, 1422 (1993)



Horst Störmer



single molecule NSOM signal

$$|\mathbf{E}(\mathbf{x}) \cdot \mathbf{p}|^2$$

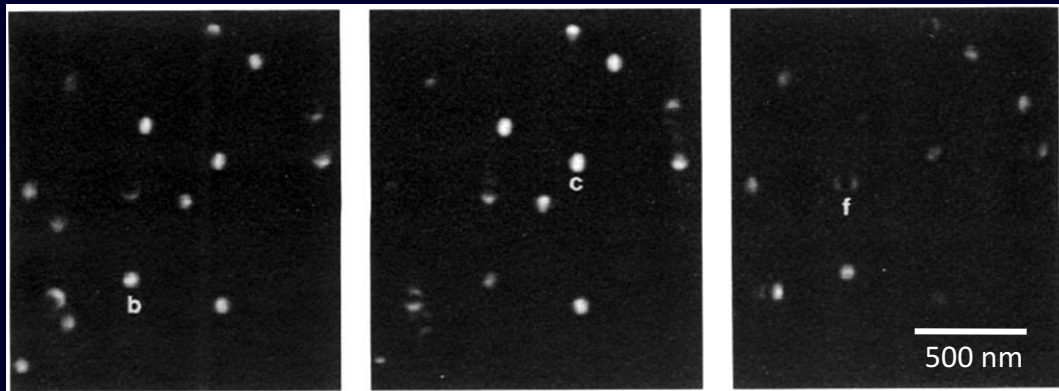
NSOM and the Birth of Single Molecule Microscopy



Rob Chichester

single molecule fluorescence anisotropy

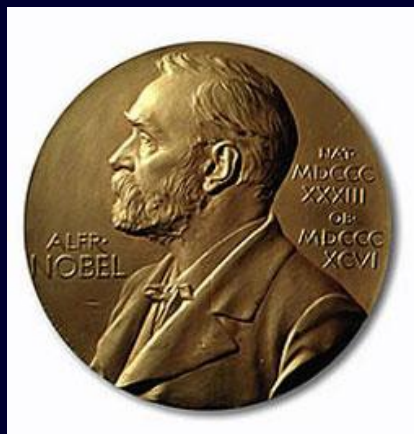
random



dil-C₁₈-(3)
molecules on
PMMA

E. Betzig, R.J. Chichester,
Science **262**, 1422 (1993)

Hans Bethe, 1967 Nobel in Physics



H.A. Bethe, *Phys. Rev.* **66**, 163 (1944)

E fields at aperture: theory vs. experiment

$z/a=0.1$ $z/a=0.2$ data $z/a=0.4$ $z/a=0.8$

$$|E_x|^2$$



$$|E_y|^2$$

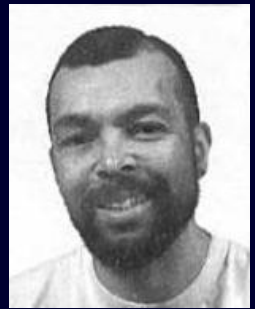


$$|E_z|^2$$



200 nm

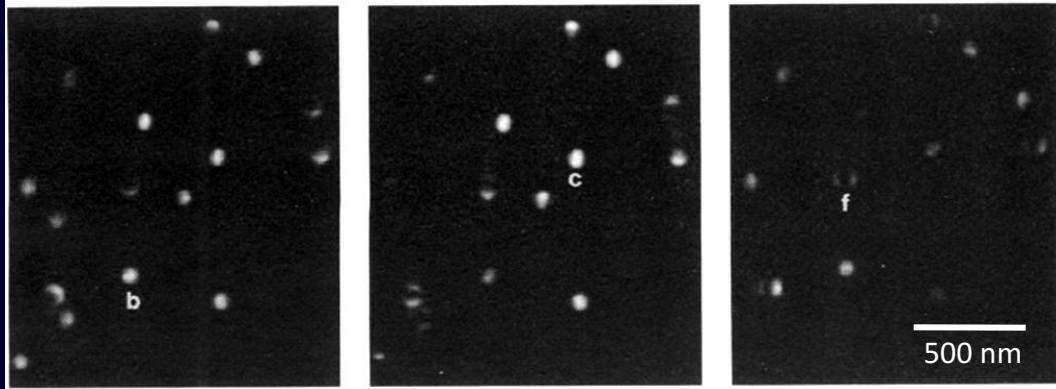
NSOM and the Birth of Single Molecule Microscopy



Rob Chichester

single molecule fluorescence anisotropy

random

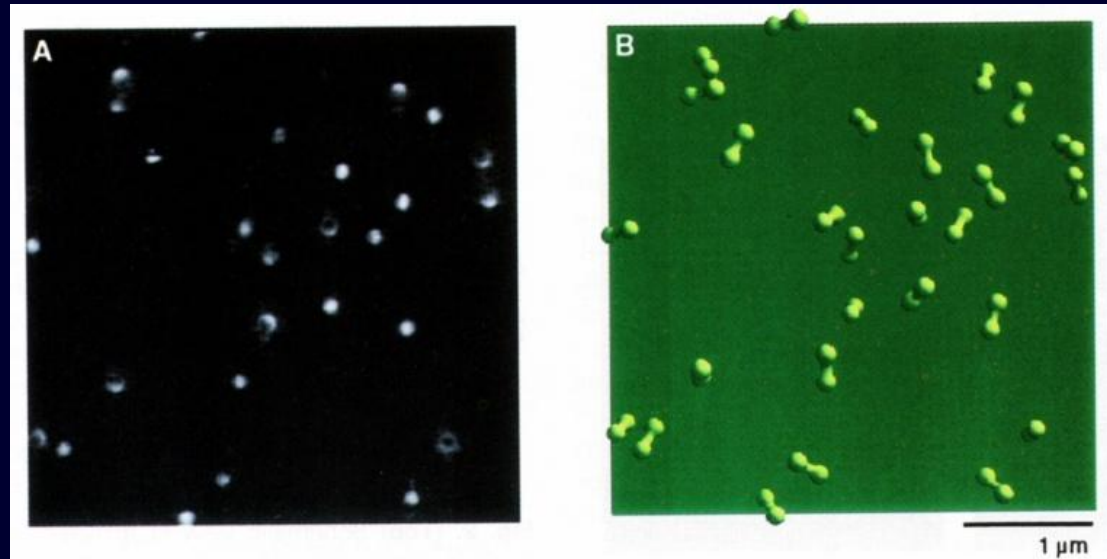


dil-C₁₈-(3)
molecules on
PMMA

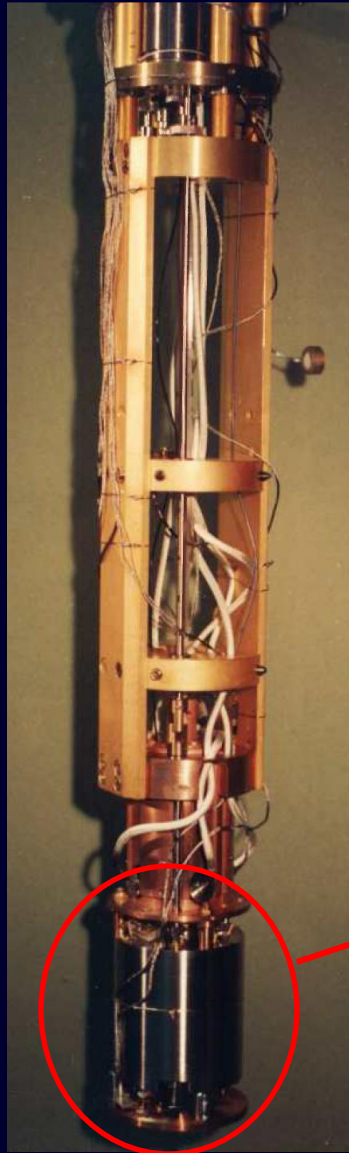
E. Betzig, R.J. Chichester,
Science **262**, 1422 (1993)

- first imaging of single molecules at room temp
- first **super-resolution** imaging of single molecules
- first measurement of single molecule dipole orientations
- first **localization** of single molecules to fraction of PSF width (12 nm xy, 6 nm z)

single molecule dipole orientations

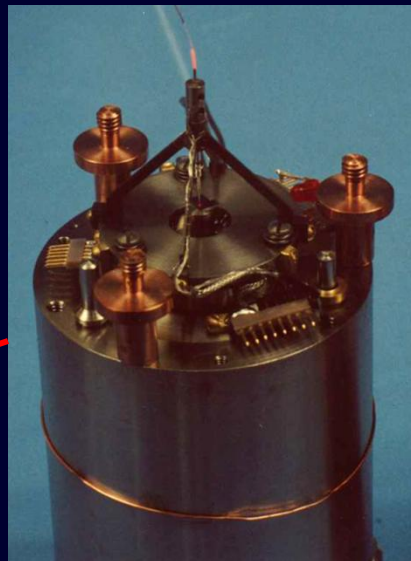


Cryogenic Near-field Spectroscopy

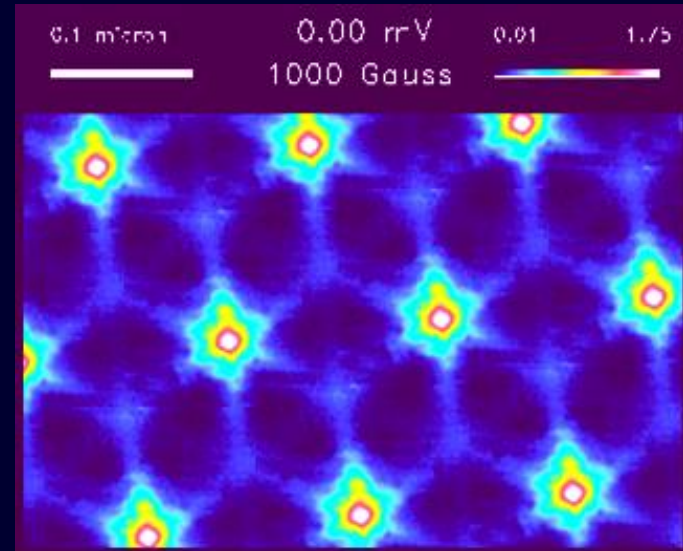


Harald Hess

Harald's low temp STM

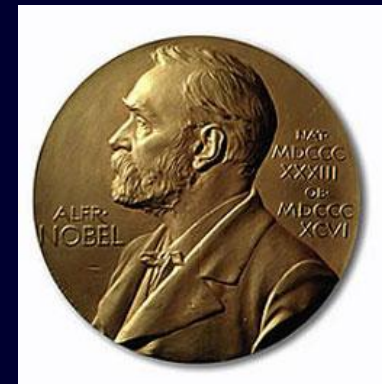


scanning tunnel spectroscopy of
Abrikosov flux lattice in NbSe₂



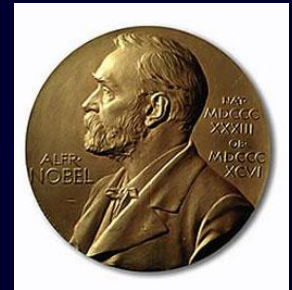
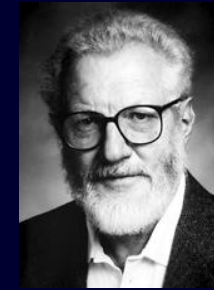
H.F. Hess, et al., *Phys. Rev. Lett.* **62**, 1691 (1989)

Alexei Abrikosov, 2003 Nobel in Physics



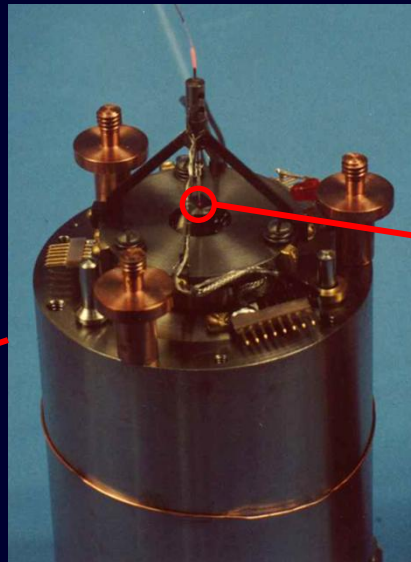
Cryogenic Near-field Spectroscopy

Alferov & Kroemer, 2000 Nobel in Physics



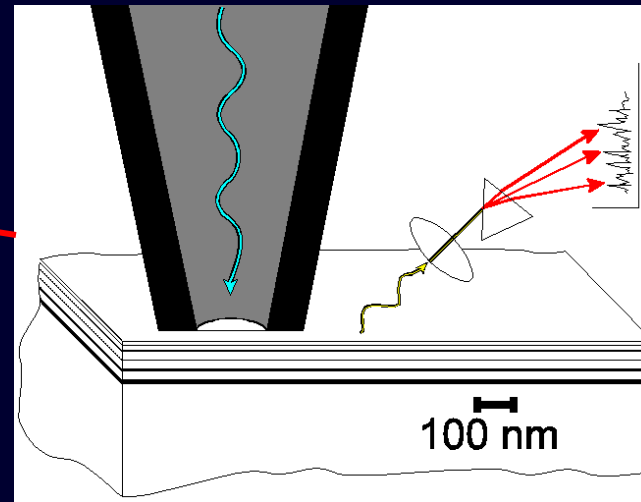
Harald Hess

Harald's low temp STM

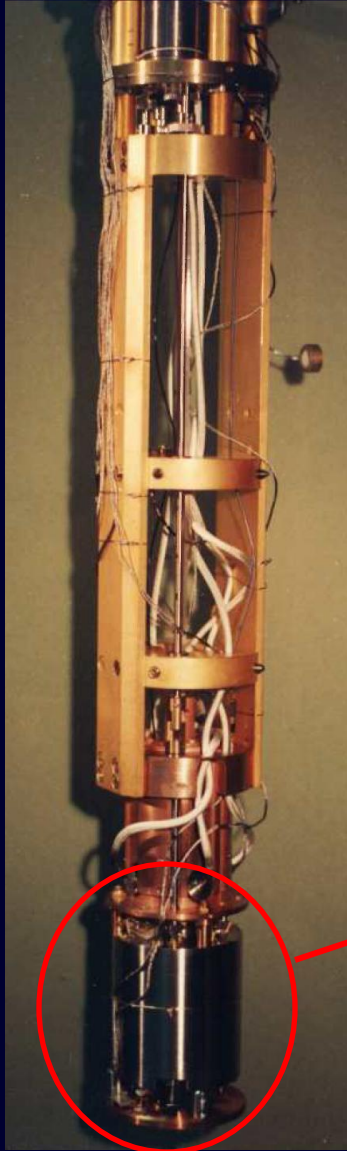


semiconductor laser diode

NSOM fiber probe

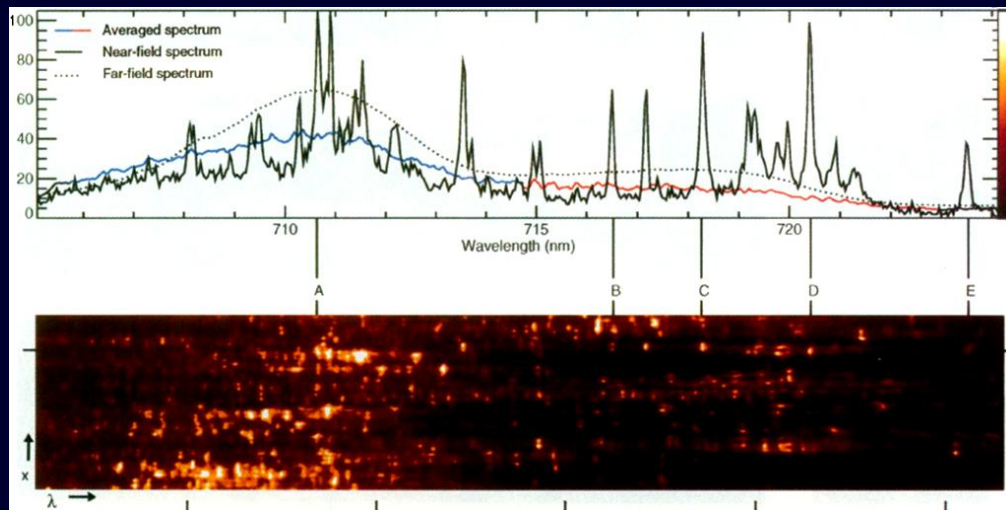


GaAs / AlGaAs multiple quantum well

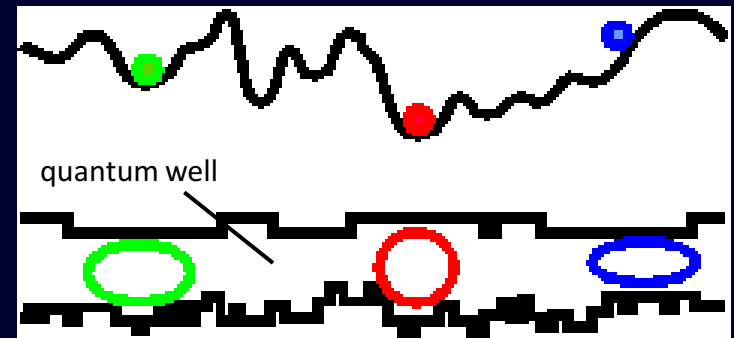


Cryogenic Near-field Spectroscopy

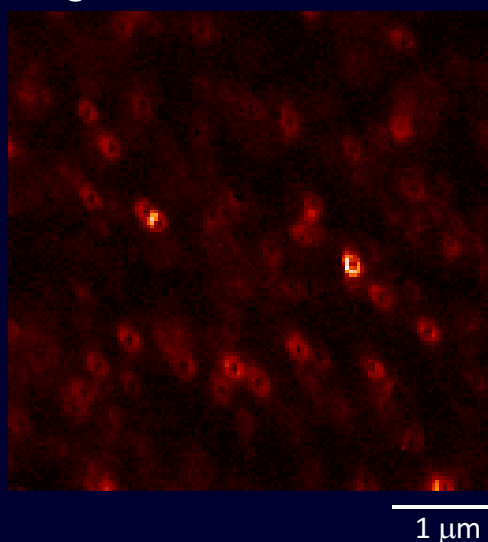
single exciton transitions, 23Å quantum well, 2°K



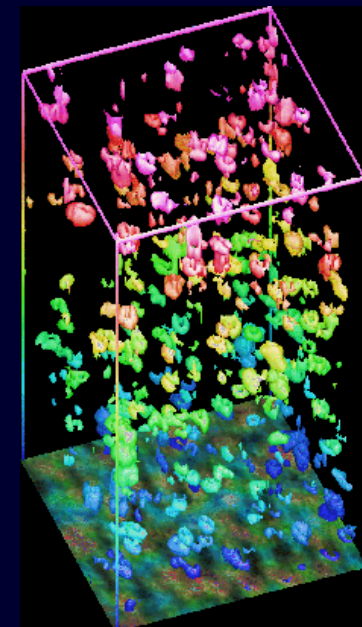
exciton energy variations due to interface roughness



exciton recombination sites
scrolling from $\lambda = 700$ to $\lambda = 730$ nm



isolation of discrete sites in x, y, λ space



H.F. Hess, E. Betzig, *et al.*,
Science **264**, 1740 (1994)

My First Mid-Life Crisis

NSOM engineering limitations:

- poor yield during manufacture
- fragile probes
- topographical artifacts
- weak signals
- probe tips get hot
- large probe tip ($0.25\ \mu\text{m}$)

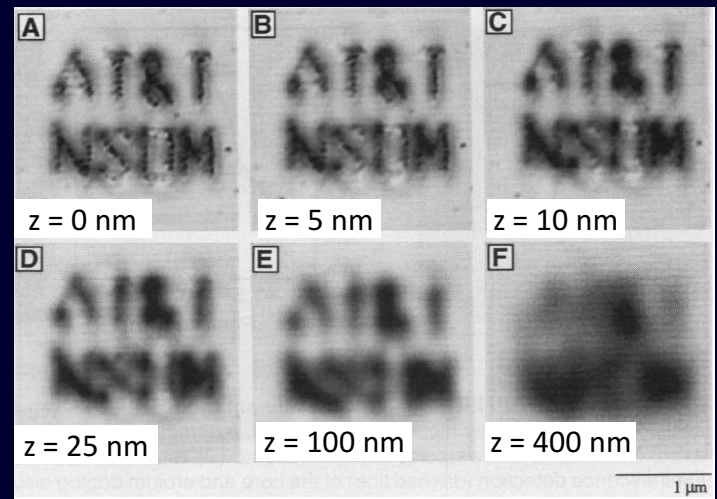
Cells aren't flat!



3D lattice light sheet microscopy,
D. Mullins, T. Ferrin, E. Betzig, *et al.*

NSOM fundamental limitations:

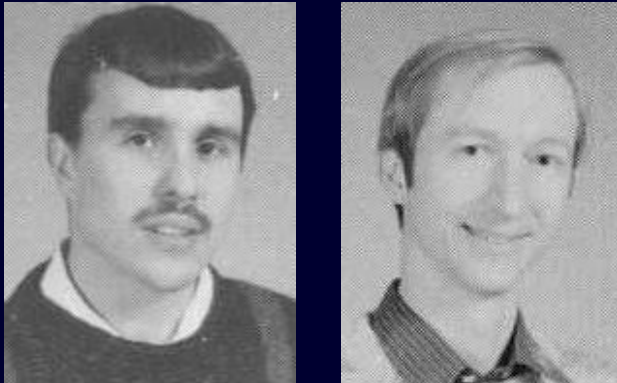
- probe perturbs fields at sample
- complex contrast mechanisms
- nonlinear image formation - artifacts
- the near-field is VERY, VERY short



E. Betzig, J.K. Trautman, *Science* **257**, 189 (1992)

My First Mid-Life Crisis

me and Harald, 1989

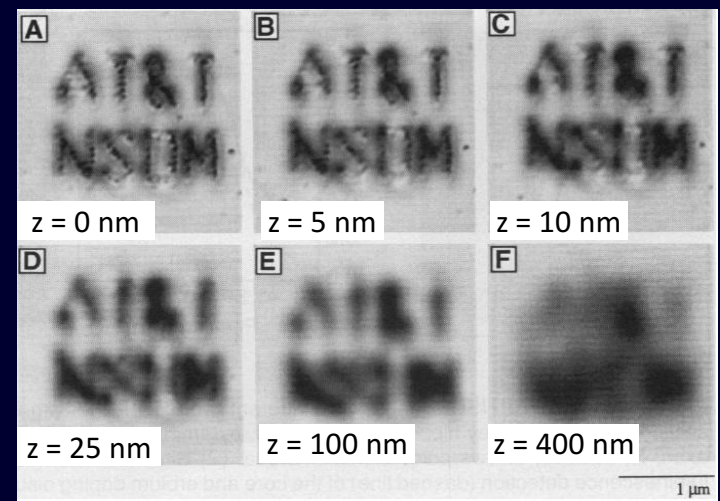


me and Harald, 1994



NSOM fundamental limitations:

- probe perturbs fields at sample
- complex contrast mechanisms
- nonlinear image formation - artifacts
- the near-field is very, very short



E. Betzig, J.K. Trautman, *Science* **257**, 189 (1992)

Multidimensional Localization Microscopy

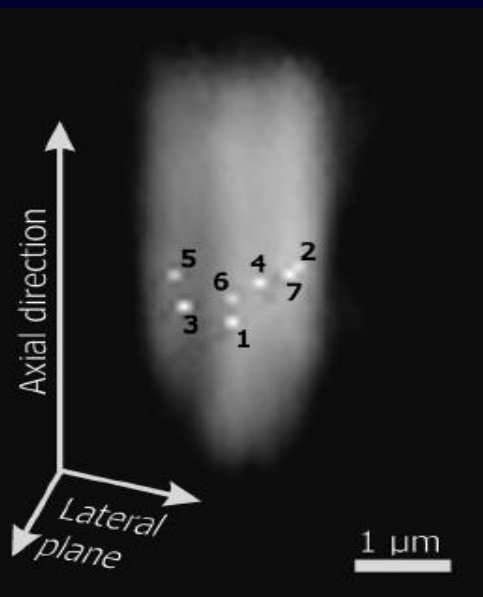
February 1, 1995 / Vol. 20, No. 3 / OPTICS LETTERS 237

Proposed method for molecular optical imaging

E. Betzig

NSOM Enterprises, 17 Webster Drive, Berkeley Heights, New Jersey 07922

spectral isolation

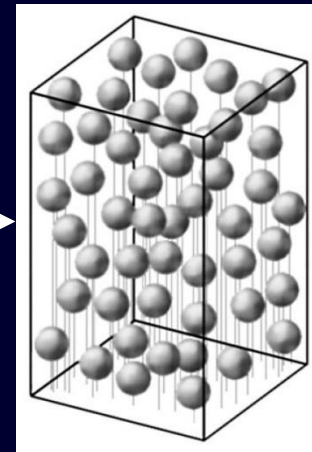


A.M. van Oijen, *et al.*, *JOSA A*16, 909 (1999)

original image



higher dimensional isolation



localization



Photobleaching: X. Qu, *et al.*, *PNAS* 101, 11298 (2004)
M.P. Gordo, *et al.*, *PNAS* 101, 6462 (2004)

Lifetime: M. Heilemann, *et al.*, *Anal. Chem.* 74, 3511 (2002)

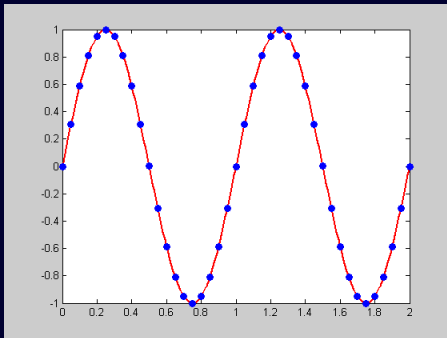
Blinking: K.A. Lidke, *et al.*, *Opt. Express* 13, 7052 (2005)

Spatial Resolution and the Nyquist Criterion

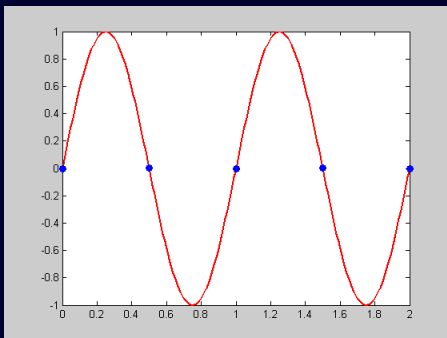
Nyquist criterion:

Sampling interval must be at least twice as fine as the desired resolution

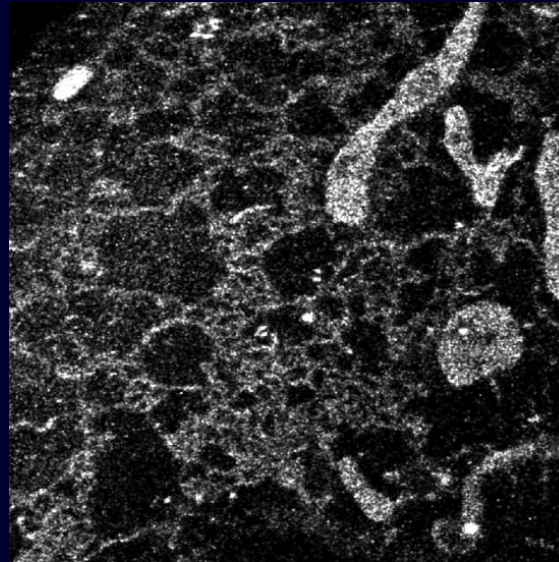
20 samples / period



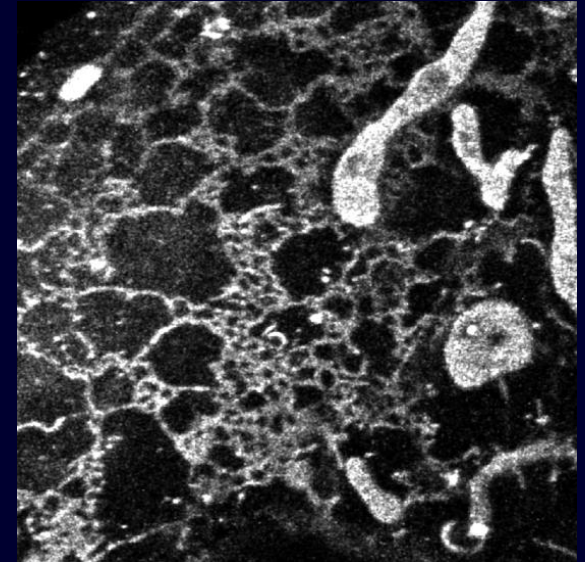
2 samples / period



initial molecular density



4× greater molecular density



2 μm

Image Dimensionality	Molecules Required per Diffraction Limited Region for 20 nm Resolution
----------------------	--

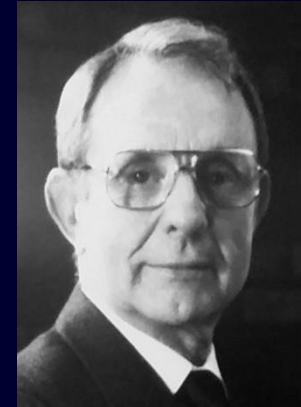
1D	25
2D	500
3D	2.9×10^4

Diffraction Limited Region:
0.25 μm dia, 0.6 μm long

And Now for Something Completely Different

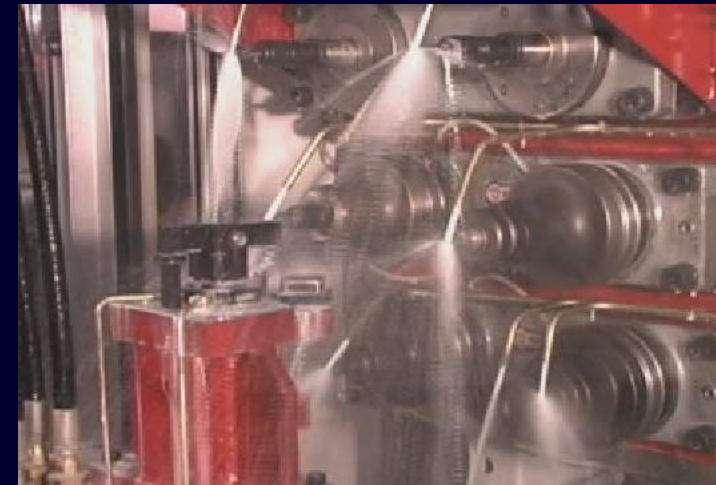


Flexible Adaptive Servohydraulic Technology (FAST)



Robert Betzig

- moves 4000 kg load at 8g acceleration
- positioning precision to 5 μm



My Second Mid-Life Crisis

Searching for a New Direction

me in Joshua Tree National Park



Harald in Sedona, Arizona



me in Oahu, Hawaii

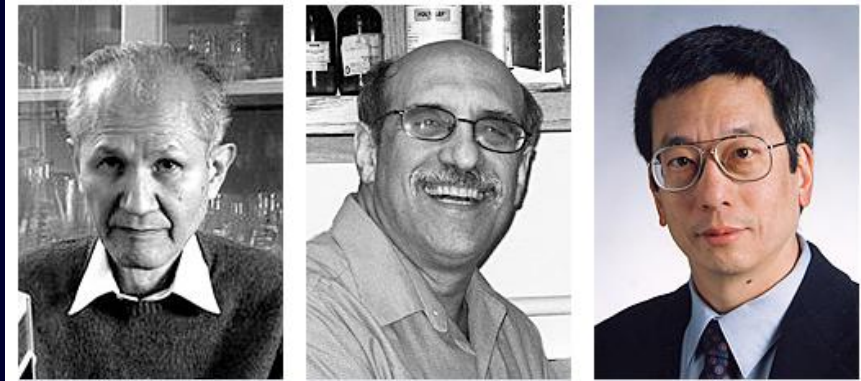


Harald in Yosemite National Park

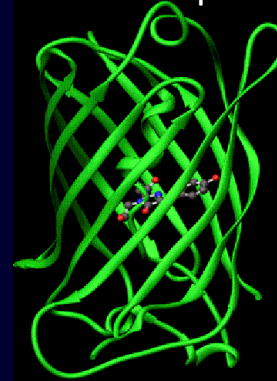


Fluorescent Proteins Revolutionize Biological Imaging

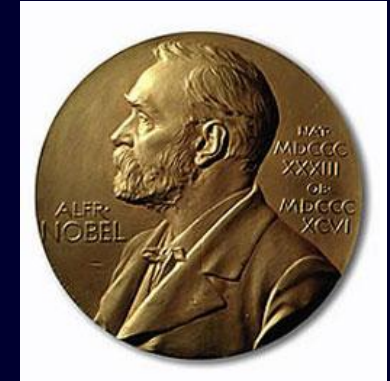
Shimomura, Chalfie, & Tsien



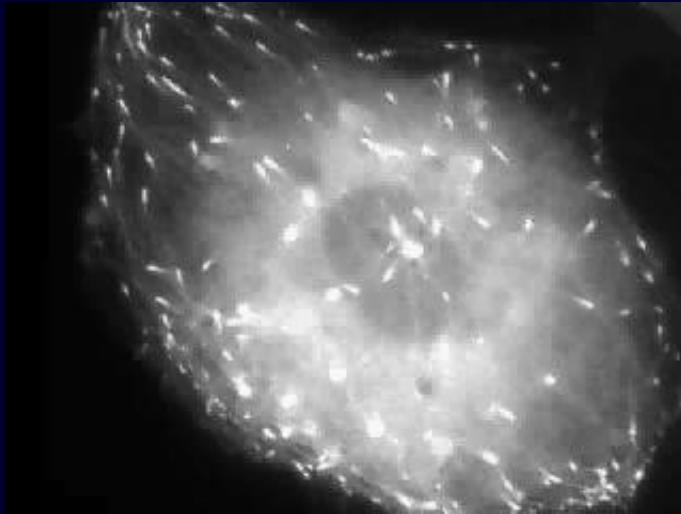
1994: green fluorescent protein



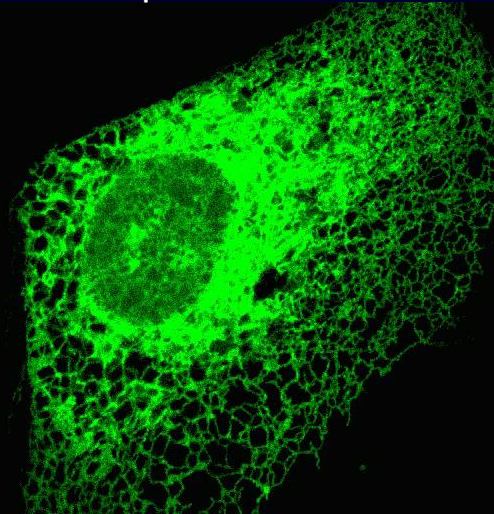
2008: Chemistry Nobel



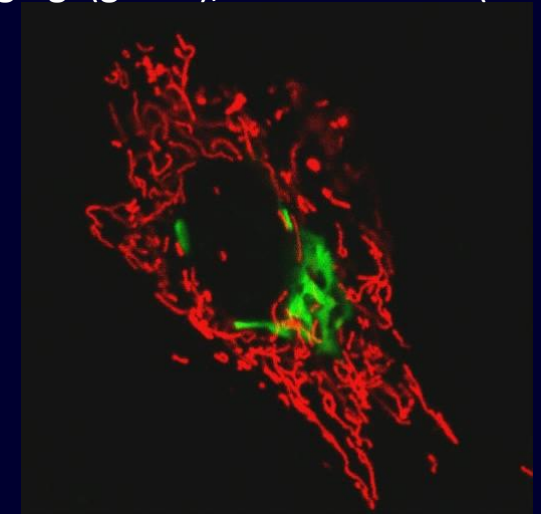
microtubule ends



endoplasmic reticulum



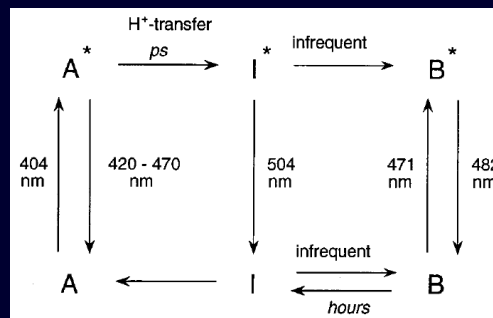
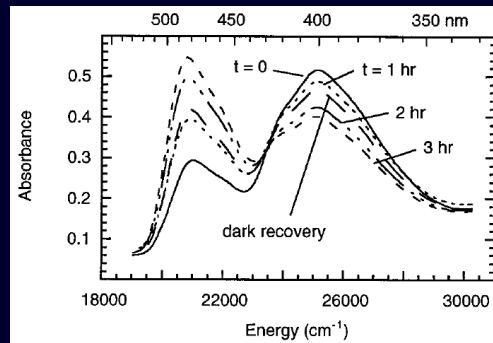
golgi (green), mitochondria (red)



Switching Behavior in Green Fluorescent Protein

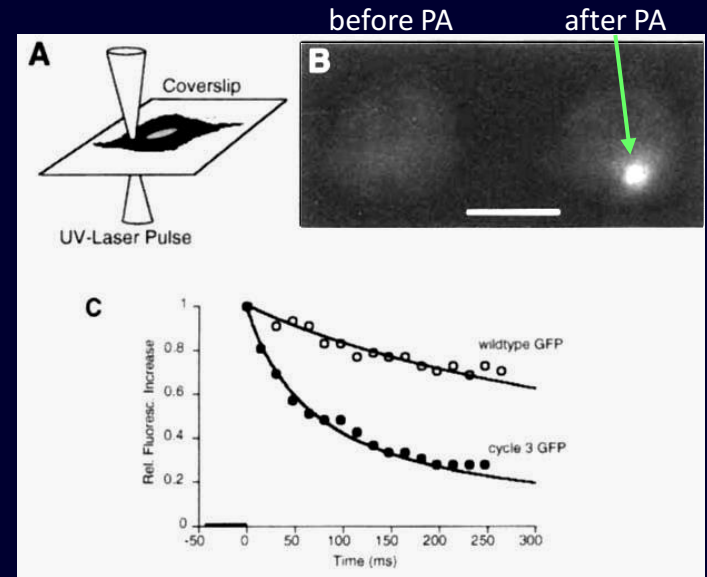
488 nm absorption increase under 398 nm illumination

proposed mechanism



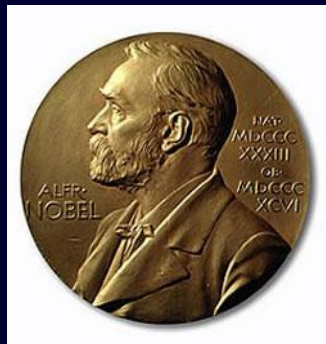
M. Chatteraj, et al., *PNAS* **93**, 8362 (1996)

in vivo UV photoactivation (PA) of wtGFP

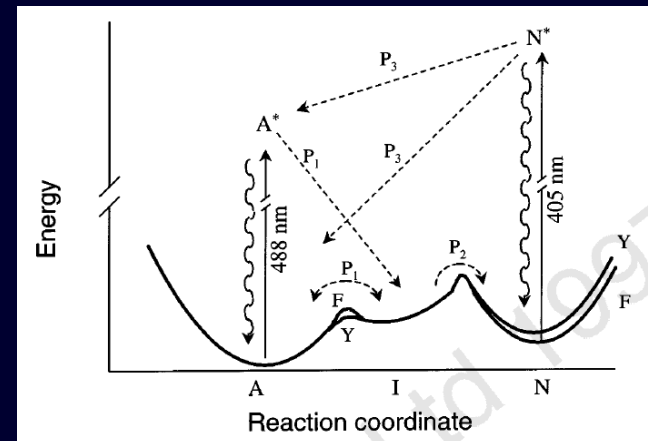


H. Yokoe, T. Meyer, *Nat. Biotech.* **14**, 909 (1996)

W.E. Moerner, 2014 Nobel in Chemistry



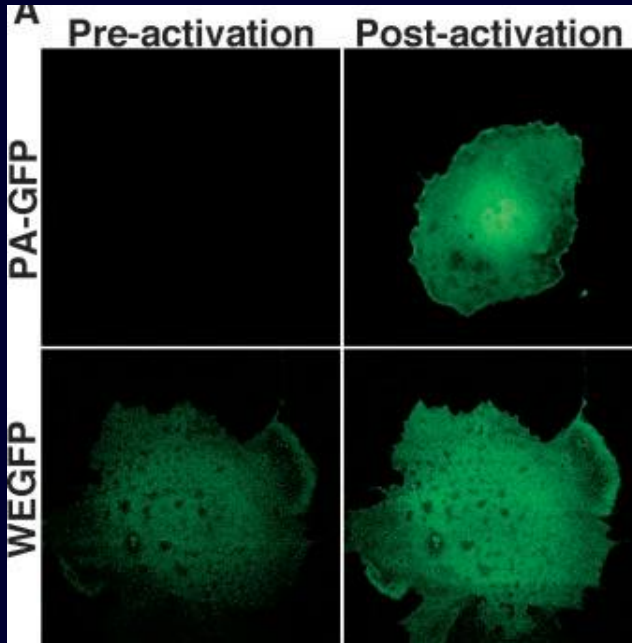
photoactivation energy diagram



R.M. Dickson, et al., *Nature* **388**, 355 (1997)

Directed Mutagenesis of Photoactivated Fluorescent Proteins (PA-FPs)

increased on/off contrast of PA-GFP



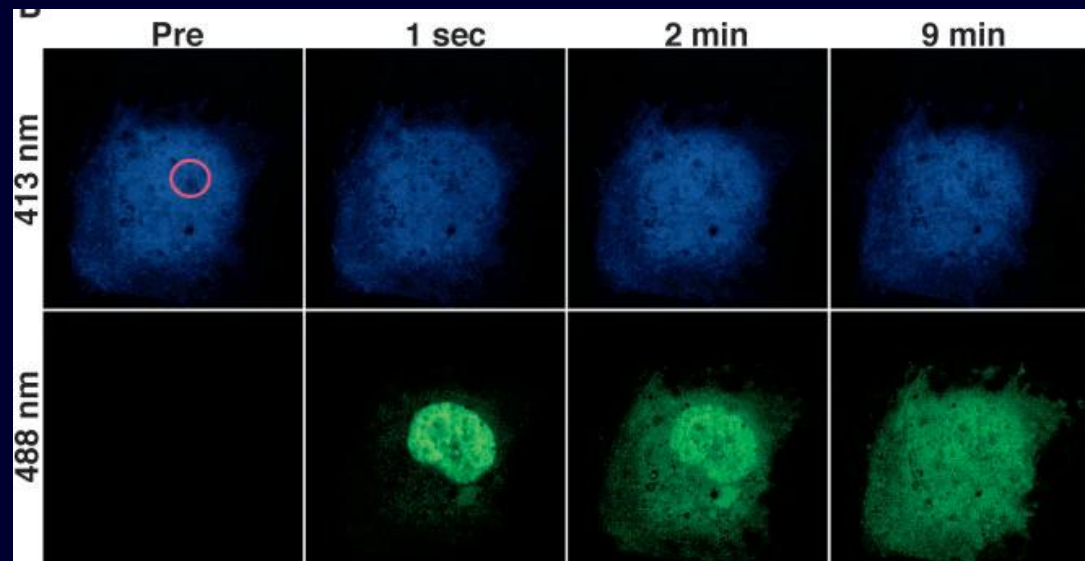
Jennifer
Lippincott-
Schwartz



George
Patterson



pulse chase: nuclear vs cytosolic diffusion



A Fateful Trip

Greg Boebinger



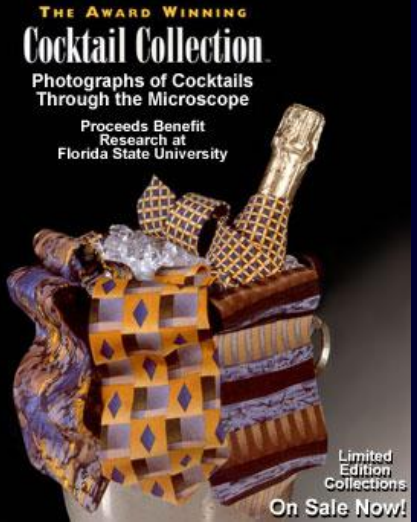
National High Magnetic Field Lab



Mike Davidson



Neckties®



Journal of Cell Science
anatomical society

Fluorescence, Florida

Gert-Jan Kremers, Sarah G. Gilbert, Paula J. Cranfill, Michael W. Davidson and David W. Piston

CHROMA

Excitation and emission spectral properties of the brightest fluorescent proteins

Protein	Excitation maximum (nm)	Emission maximum (nm)	Extinction coefficient (x10 ⁴ M ⁻¹ cm ⁻¹)	Quantum yield	Relative brightness (% of EGFP)
mTagBFP	309	456	82.0	0.63	98
mCherry	610	633	120.0	0.64	75
mEGFP	488	507	96.0	0.60	100
mVenus	515	528	82.2	0.57	106
mKO2	551	569	82.8	0.62	118
mApple	568	592	75.0	0.48	109
mCherry	587	610	72.0	0.22	47
mKate2	588	633	82.0	0.40	74

Fluorescent protein properties

Protein	Excitation maximum (nm)	Emission maximum (nm)	Extinction coefficient (x10 ⁴ M ⁻¹ cm ⁻¹)	Quantum yield	Relative brightness (% of EGFP)
mTagBFP	309	456	82.0	0.63	98
mCherry	610	633	120.0	0.64	75
mEGFP	488	507	96.0	0.60	100
mVenus	515	528	82.2	0.57	106
mKO2	551	569	82.8	0.62	118
mApple	568	592	75.0	0.48	109
mCherry	587	610	72.0	0.22	47
mKate2	588	633	82.0	0.40	74

β-barrel motif

β-barrel motif structure showing N-terminus, C-terminus, β-barrel, Chromophore, β-sheet, Loop, and ~3 nm scale.

Critical mutations

Mutation	Properties
S205F	Increases folding rate, enhances protein stability
F66L	Accelerates chromosome formation
D98M	Improves stability and pH resistance, photostability and folding
S73A	Faster folding rate, stabilizes protein
S147P	Faster maturation rate, located near chromophore
R156K	Faster folding rate, stabilizes protein
V163A	Reduces hydrophobicity, no effect on folding rate
F167T	Reduced fluorescence, faster maturation rate

Fluorescent protein localization

Protein	Localization
mTagBFP	Microtubules
mCherry	Intermediate filaments
mKO2	Actin filaments
mCherry	Plasma membrane
mCherry	Mitochondria plus other
mCherry	Lysosomes
mCherry	Peroxisomes
mCherry	Tight junctions
mCherry	Single vesicles
mCherry	Nuclear envelope
mCherry	Auto phagosomes
mCherry	Cellular lysosomes
mCherry	Cytoskeleton filaments
mCherry	Cytoskeleton
mCherry	Cellular organelles
mCherry	Endoplasmic reticulum

Multi-color imaging using fluorescent protein fusions

LLC-PK1 (pig kidney) cells expressing mTagBFP fused to a tubulin (green) and mApple fused to histone H2B (red).

HeLa (human embryonic) cells expressing mCherry fused to a Golgi targeting peptide (green), mCherry fused to a nucleus targeting signal (yellow), and mCherry fused to a mitochondria-targeting peptide (red).

HeLa (human embryonic) cells expressing mTagBFP fused to histone H2B (blue), mTagBFP fused to peroxisomal membrane protein (cyan), mEGFP fused to Listeria (cyan, green), and mCherry fused to zymosan (red, yellow, purple, and mKate2 fused to peroxisome dehydrogenase (mitochondria, red).

HeLa cells expressing mTagBFP fused to histone H2B (blue), mTagBFP fused to peroxisomal membrane protein (cyan), mEGFP fused to galactosyl 6 α -mannose (cyan, green), mKO2 fused to zymosan (red, yellow, purple, and mKate2 fused to peroxisome dehydrogenase (mitochondria, red).

Abbreviations: C-CAA, C-terminal amino acid sequence for fatty acylation; CENPB, centromere protein B; C-42, cytoskeleton 42; EGFP, enhanced green fluorescent protein; G, Golgi; H2B, histone H2B; K, Katushika orange fluorescent protein; LAMP1, lysosomal membrane glycoprotein 1; C-C, cytoskeleton high-chain stabilizer; MTS, mitochondria targeting signal; VE, vesicle; vesicular epithelial marker.

Fluorescent Protein Source References: mApple: Shaner, N. C. et al. (2008). Nat. Methods 5, 685-689; mCherry: Shaner, N. C. et al. (2005). Nat. Methods 2, 1016-1022; mEGFP: Cormack, B. P. et al. (1996). Gene 173, 53-58; mKate2: Shcherbo, D. et al. (2007). Science 316, 907-910; mKO2: Shaner, N. C. et al. (2008). Chem Biol 16, 132-140; mTagBFP: Subram, C. M. et al. (2008). Chem Biol 16, 1119-1124; mTagBFP: Subram, C. M. et al. Nat. Methods 7, 137-139; mVenus: Nagai, T. et al. (2002). Nat.

website tutorials

Zeiss



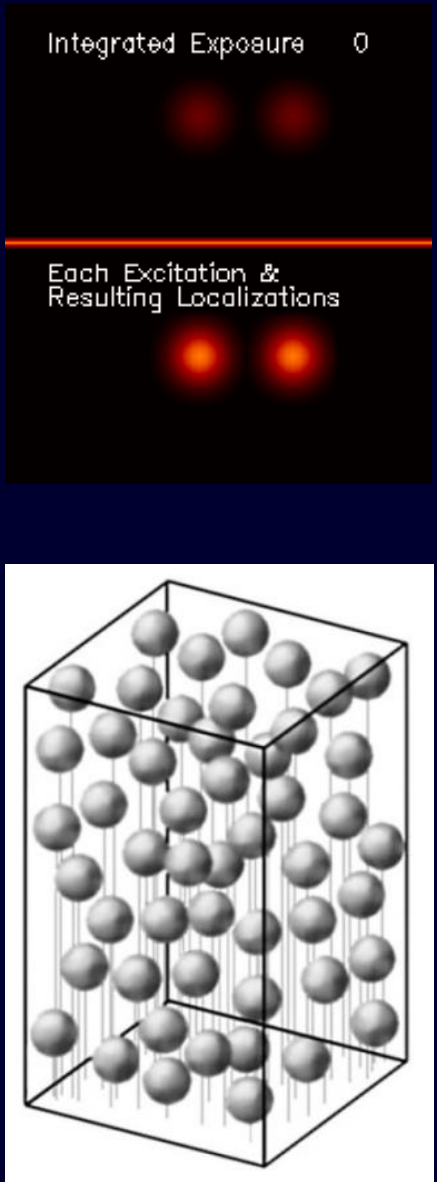
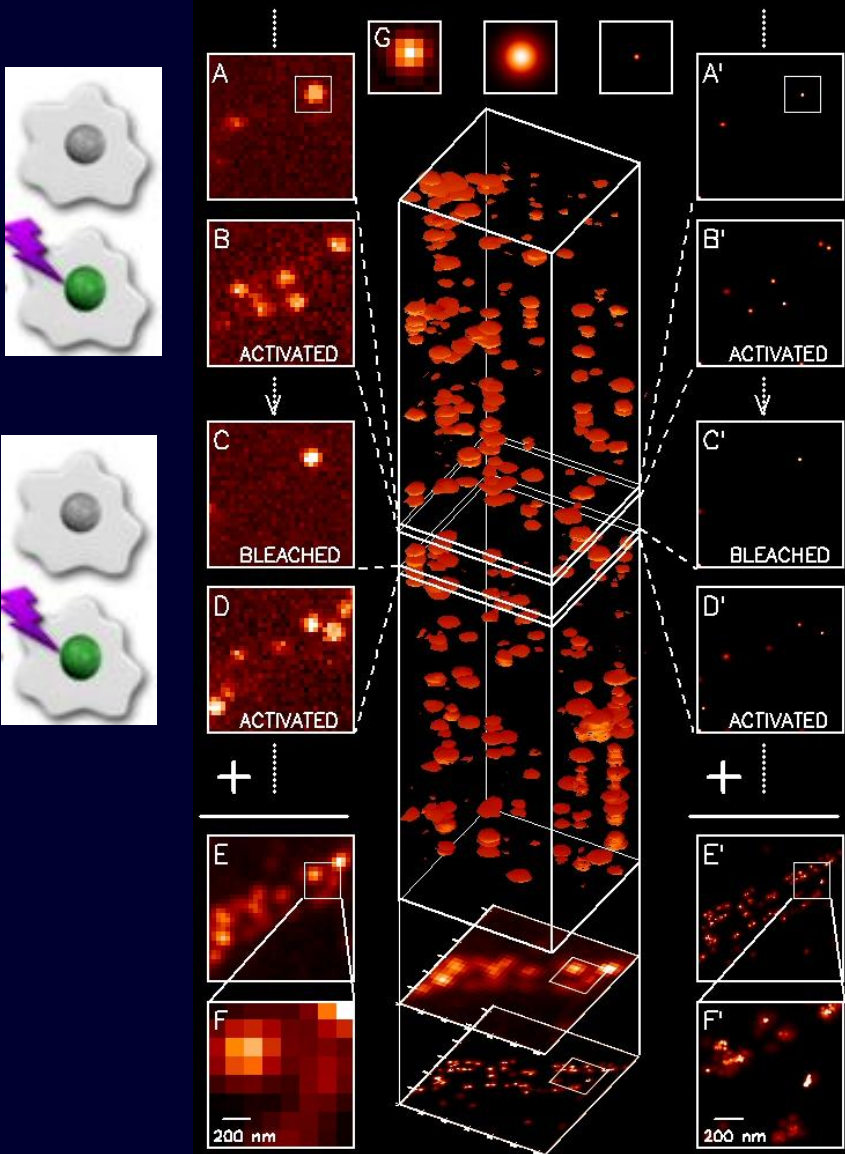
Olympus



Nikon

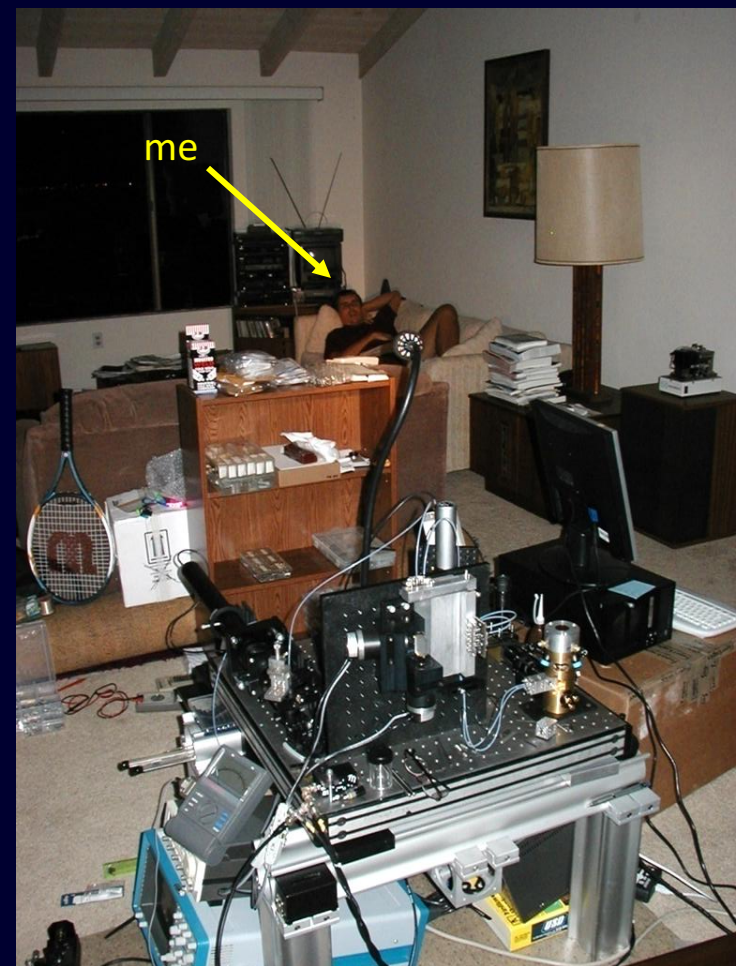


Finding the Missing Link



E. Betzig, *et al.*, *Science* **313**, 1642 (2006)

La Jolla Labs



Assembling the Rest of the Team

Jennifer
Lippincott-
Schwartz

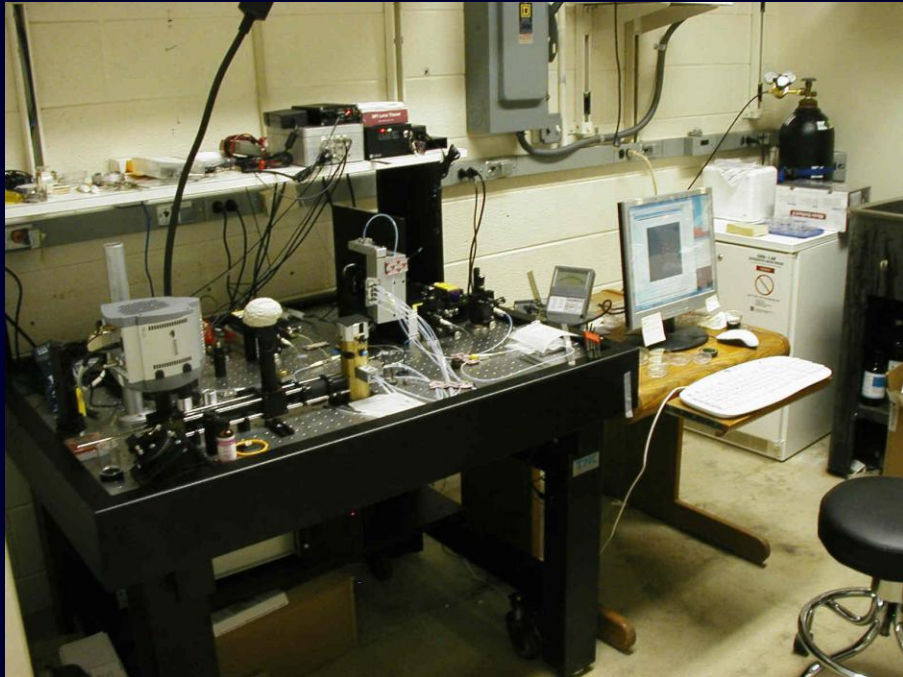


George
Patterson



Rob Tycko, NIDDK

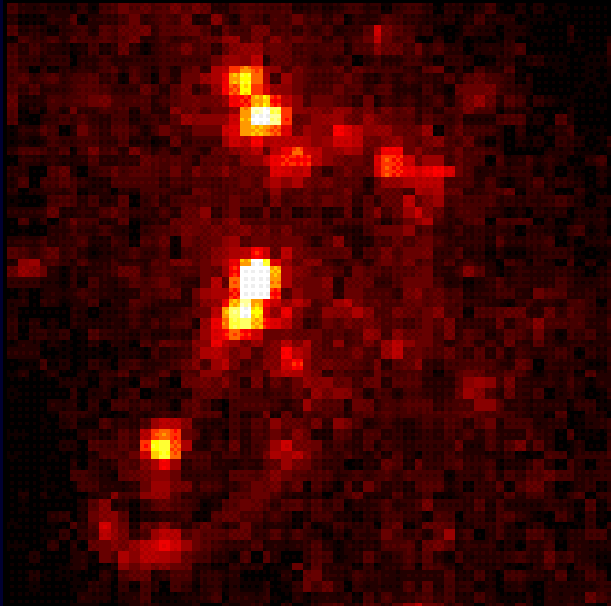
the microscope in the darkroom in Jennifer's lab



Photoactivated Localization Microscopy (PALM)

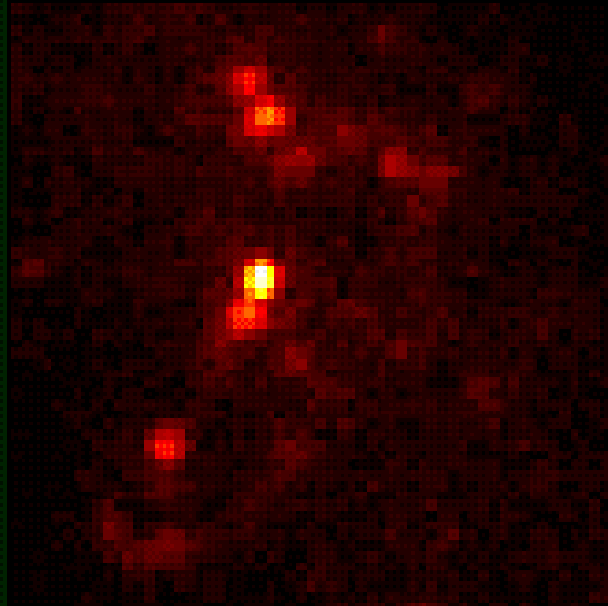
lysosomes, COS-7 cell, Kaede-tagged CD63

single molecule frames

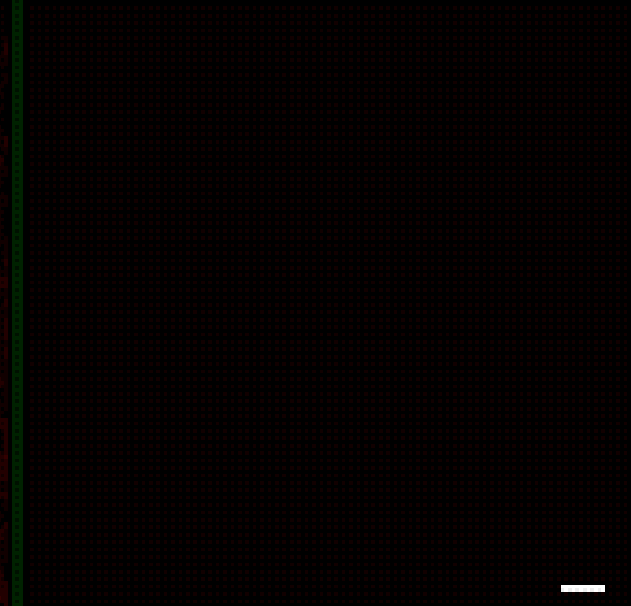


0.5 sec/frame

integrated image



PALM image



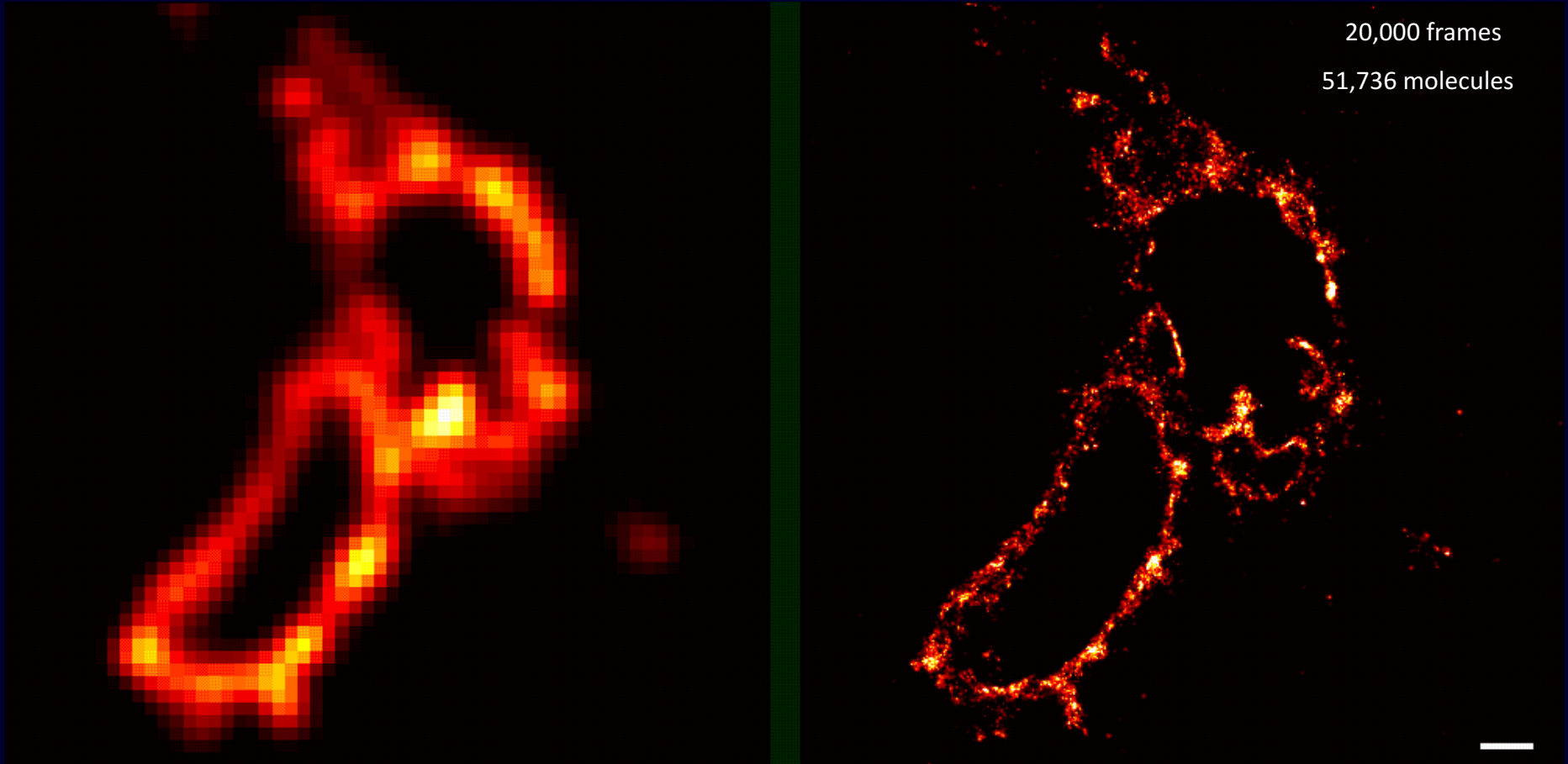
0.5 μm

- ~80 nm cryosection:
- low autofluorescence
 - immobile PA-FPs
 - image internal organelles

Photoactivated Localization Microscopy (PALM)

TIRF

PALM



lysosomes, COS-7 cell, Kaede-tagged CD63

0.5 μm

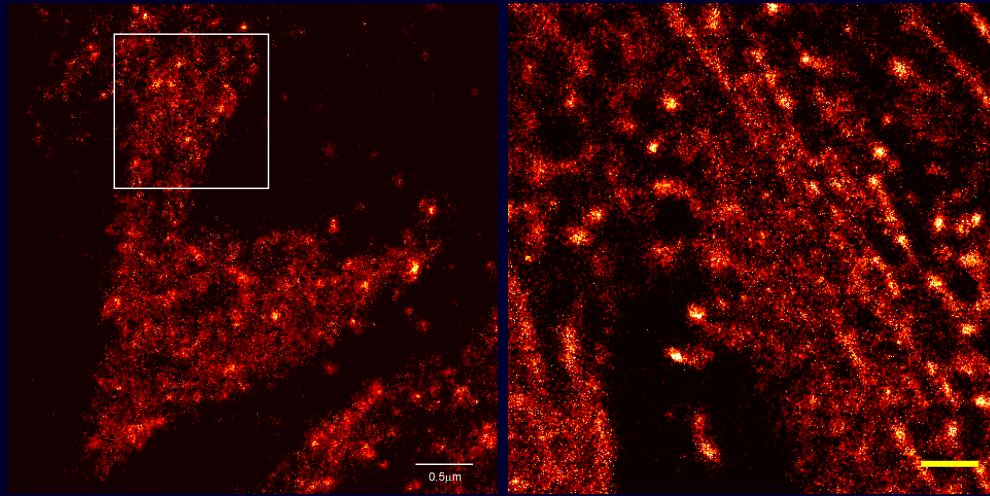
E. Betzig, *et al.*, *Science* 313, 1642 (2006)

A High On/Off Contrast Ratio is Essential for High Resolution

paxillin, focal adhesions

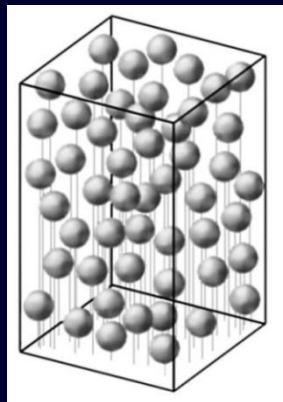
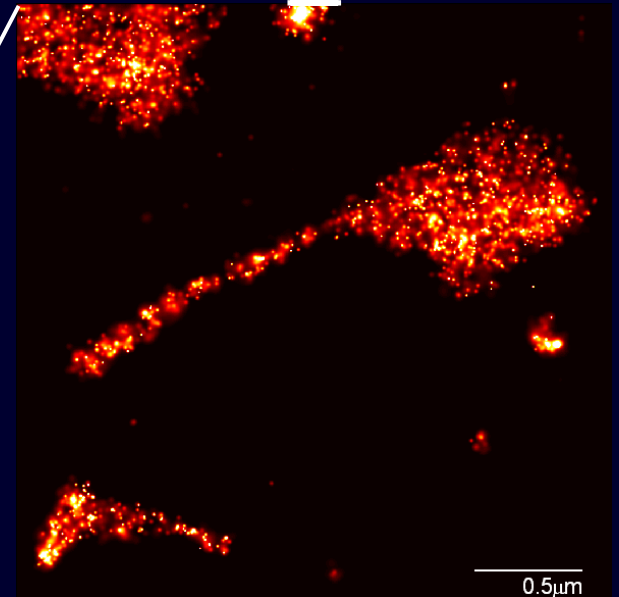
EosFP > 2000:1

PA-GFP < 75:1

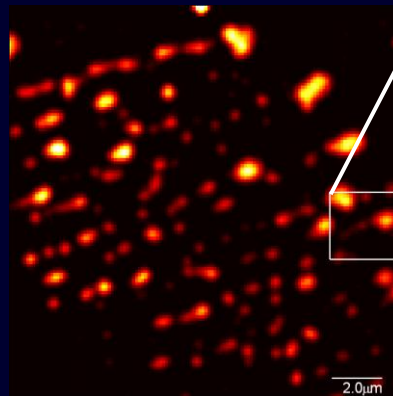


Eos FP and caged Q-rhodamine support Nyquist-defined sub-20 nm resolution

caged Q-rhodamine, > 1000:1



diffraction limited TIRF



E. Betzig, *et al.*, *Science*
313, 1642 (2006)

From Rags to Riches, Thanks to HHMI

Janelia Research Campus



The Boss: Gerry Rubin



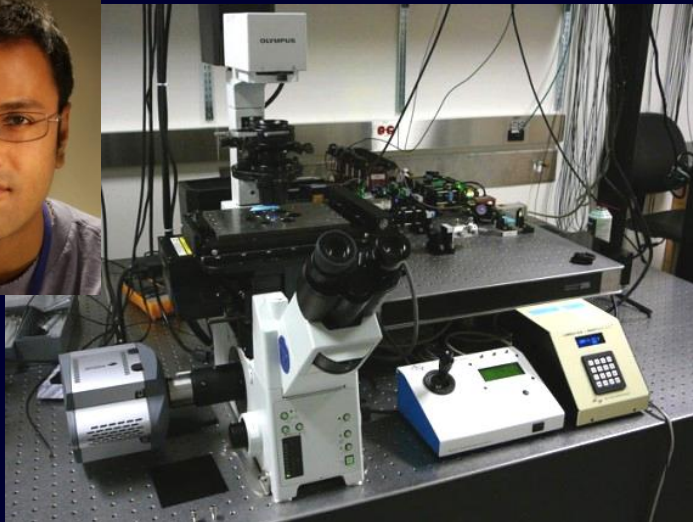
Endless Coffee



Hari Shroff



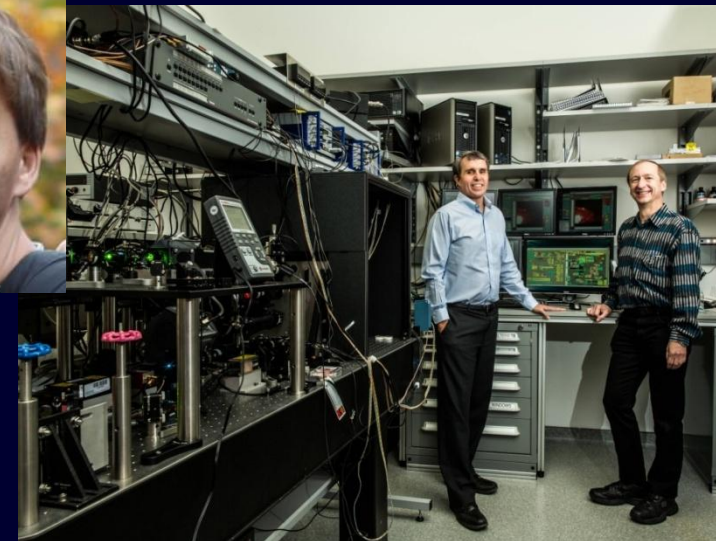
my PALM



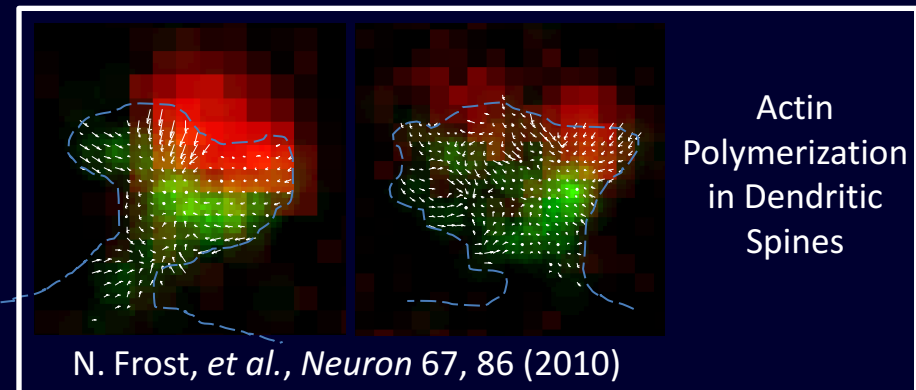
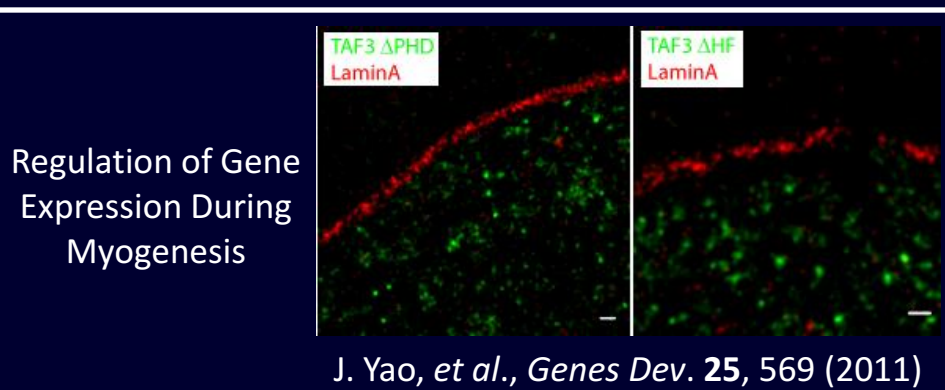
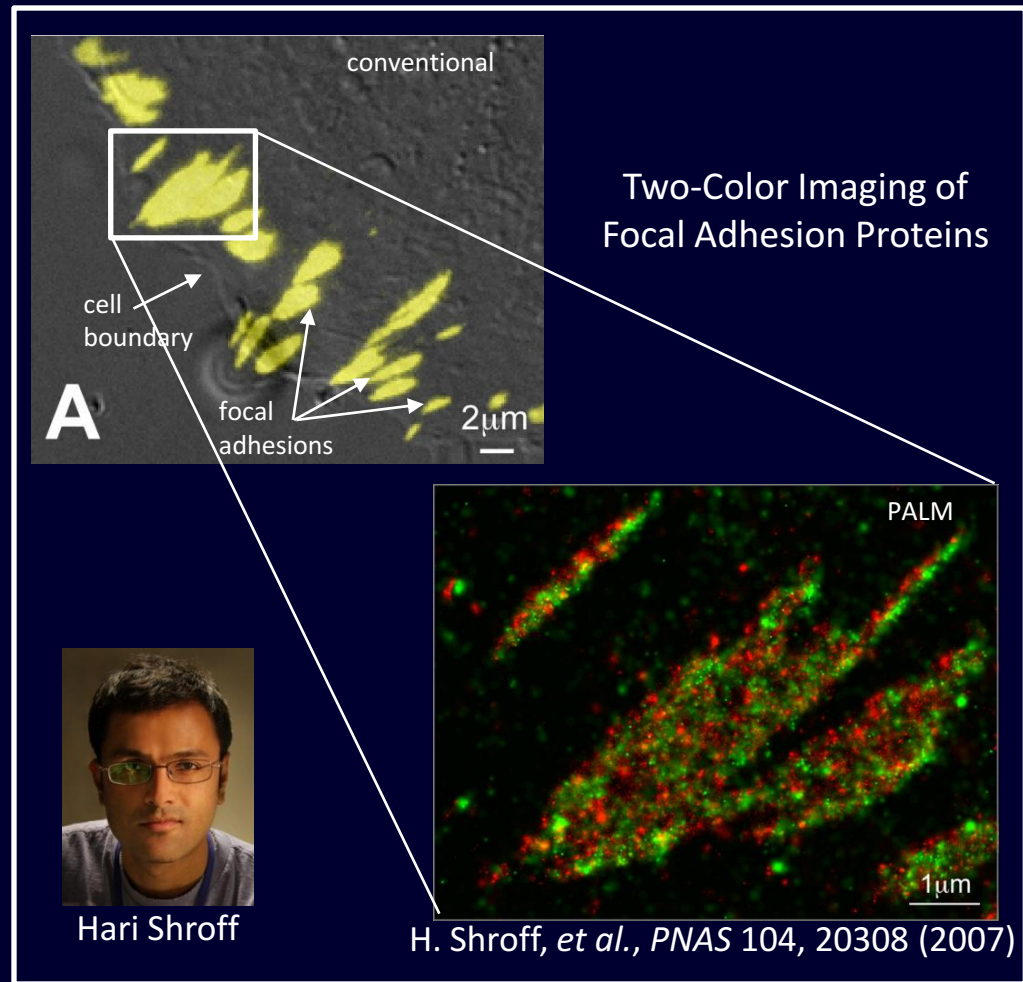
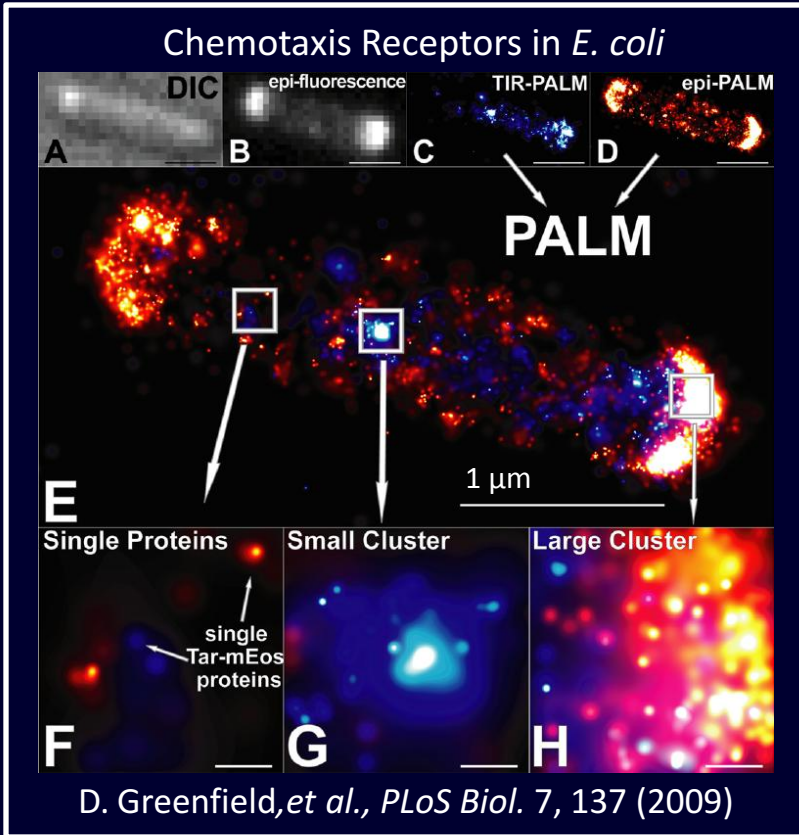
Gleb Shtengel



Harald's iPALM



PALM Application Examples



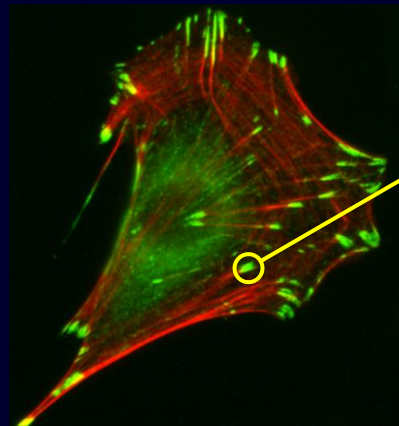
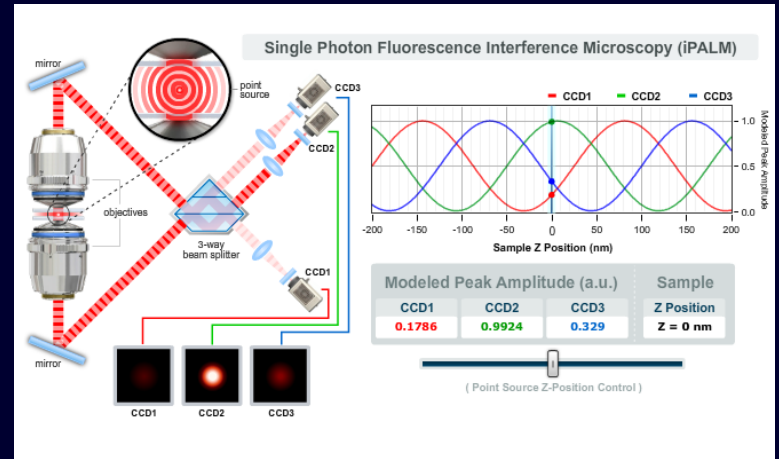
iPALM: Ultrasensitive PALM in 3D

Harald Hess



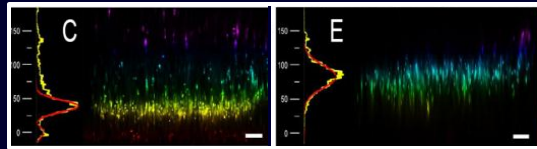
iPALM schematic

three phase single molecule interferometry

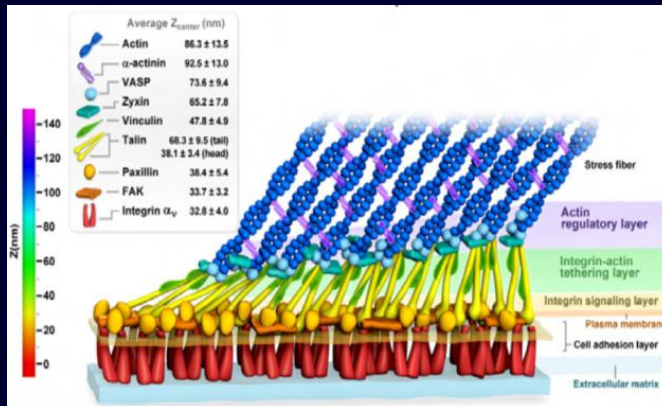


single focal adhesion

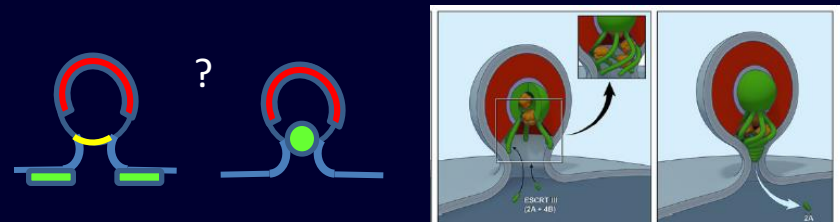
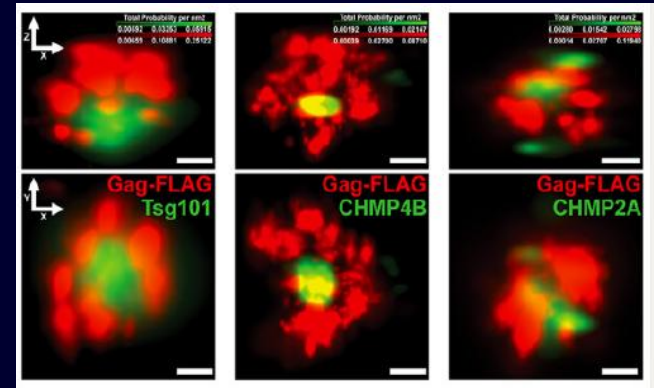
iPALM xz view



vertical architecture of adhesions



ESCRT machinery at HIV budding sites

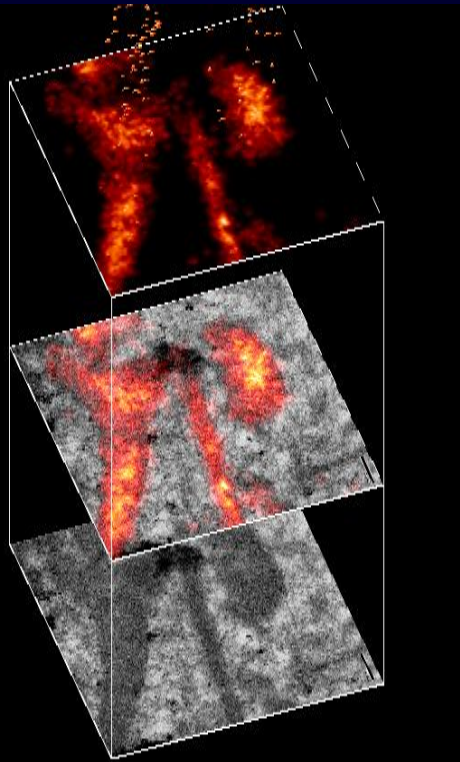


P. Kanchanawong, et al., *Nature* **468**, 580 (2010)

S.B. Van Engelenburg, et al., *Science* **343**, 653 (2014)

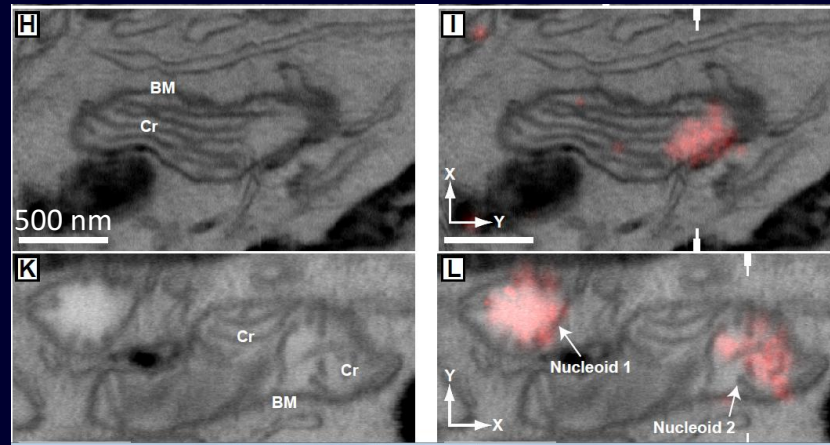
Correlative Electron Microscopy and PALM

first correlative EM
with super-resolution:
mitochondria

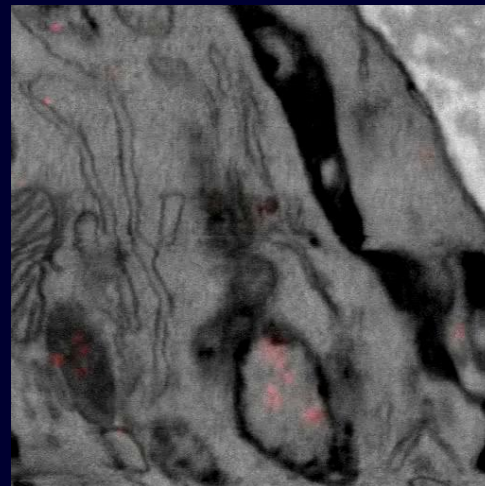


E. Betzig, *et al.*, *Science*
313, 1642 (2006)

3D correlative EM/PALM
mitochondria (B&W – FIB SEM)
mitochondrial DNA (red - iPALM)

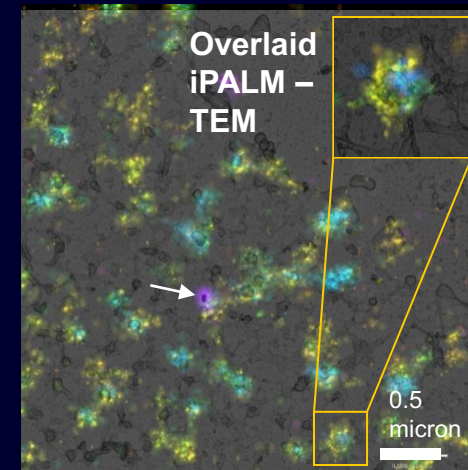
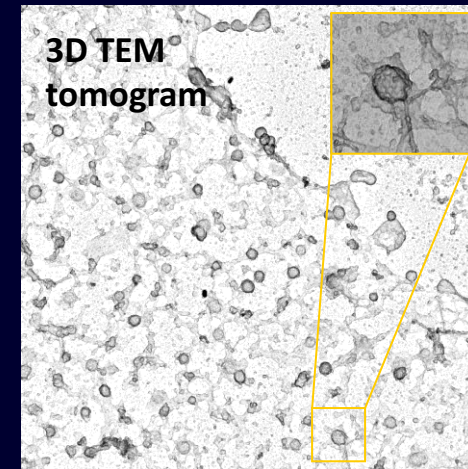


scrolling plane-by-plane thru 3D



B.G. Kopek, *et al.*, *PNAS*, **109**, 6136 (2012)

cell membrane (B&W - TEM)
& clathrin (color - iPALM)

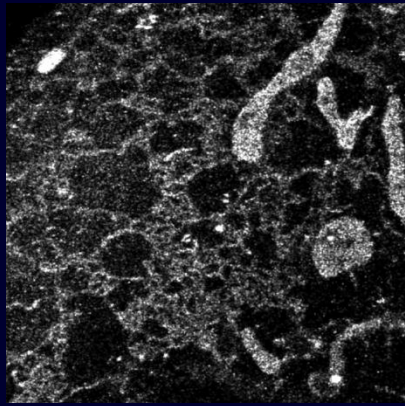


K. Sochaki, *et. al*, *Nat. Methods*, **11** 305 (2014)

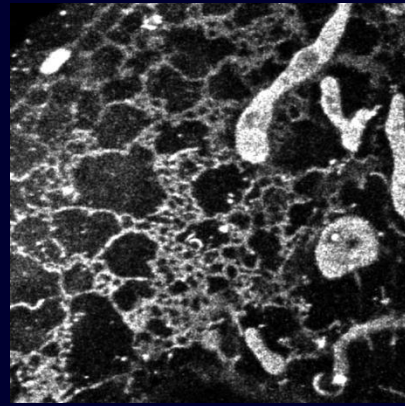
Caveats with Super-Resolution Microscopy: Fixed Cells

extremely high labeling densities required

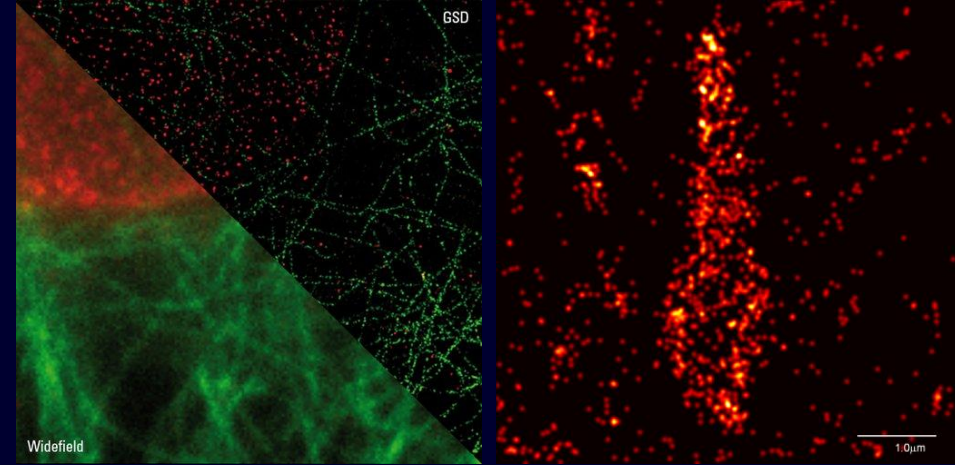
initial density



4x higher density

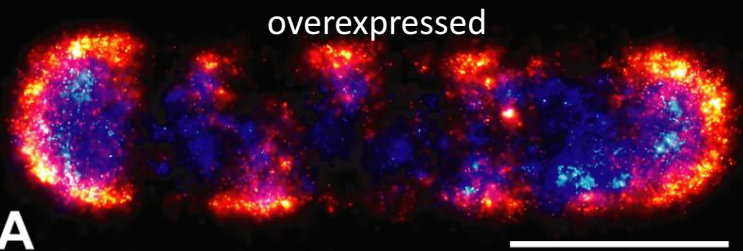


exogenous dyes: limited affinity & high background

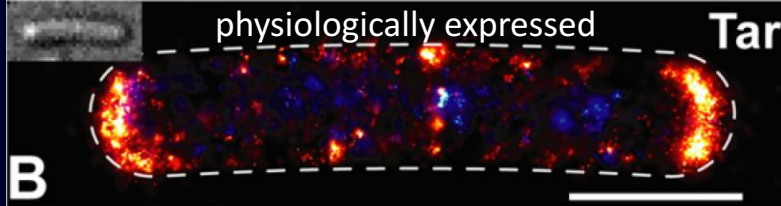


overexpression of protein

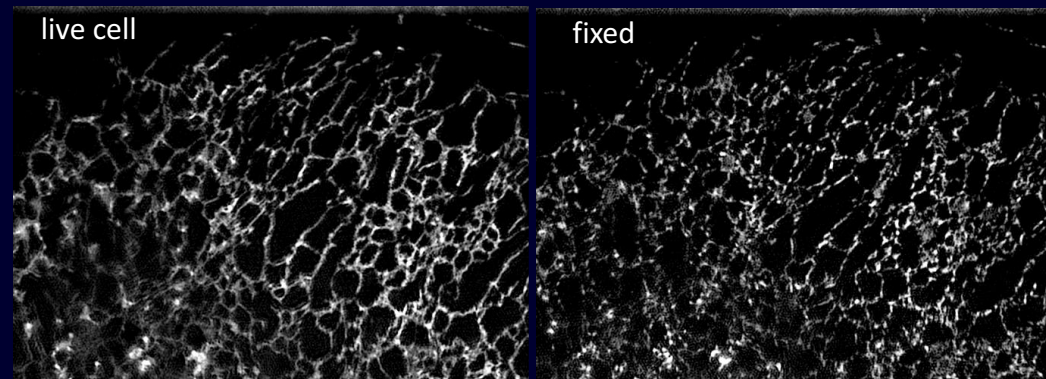
overexpressed



physiologically expressed

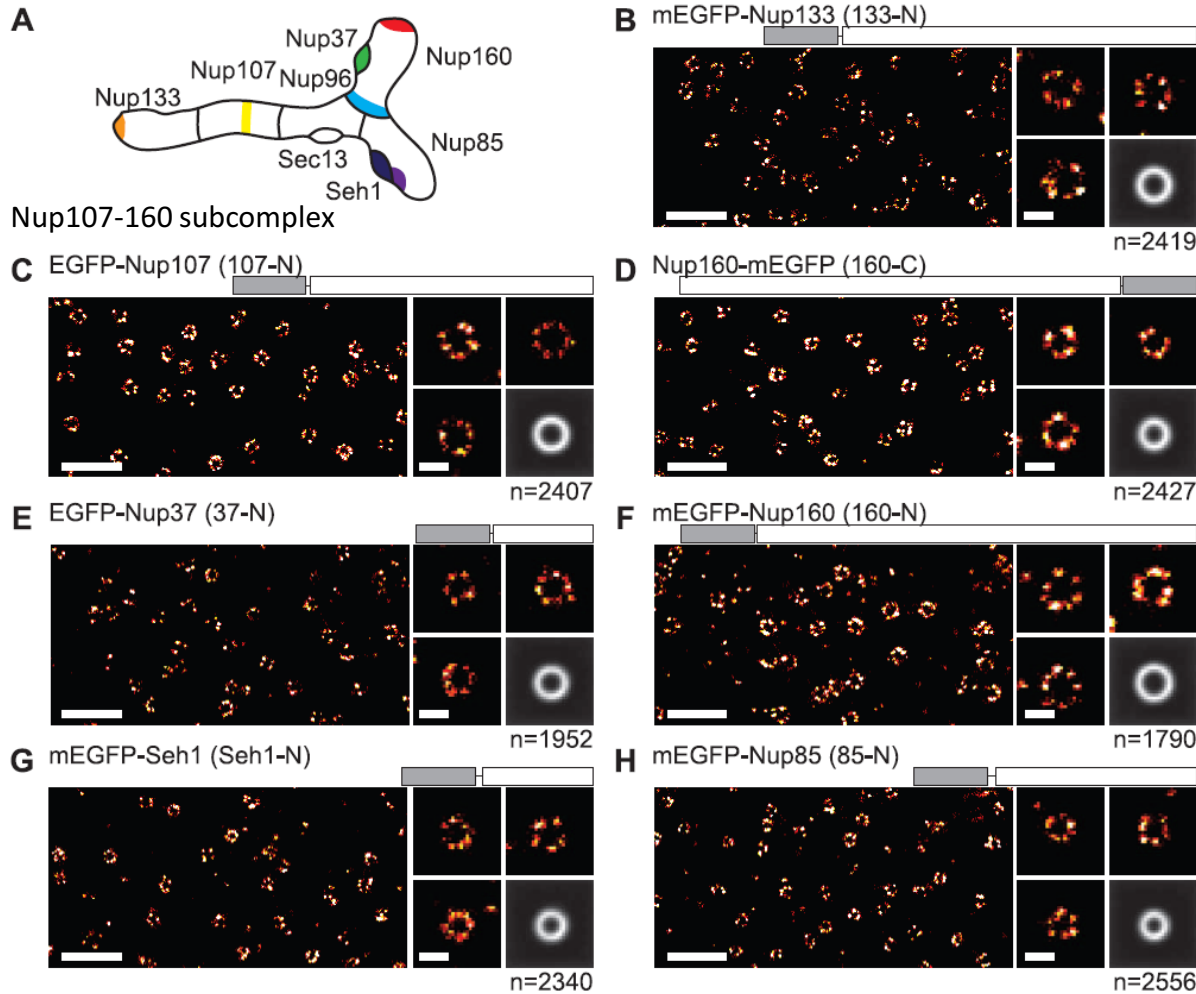


fixation artifacts, endoplasmic reticulum

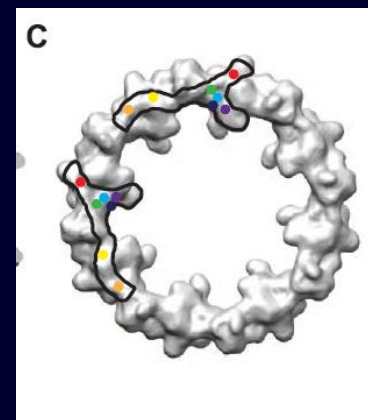
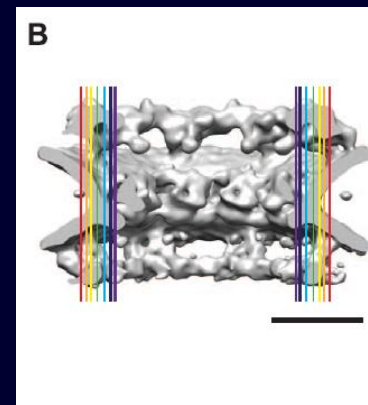
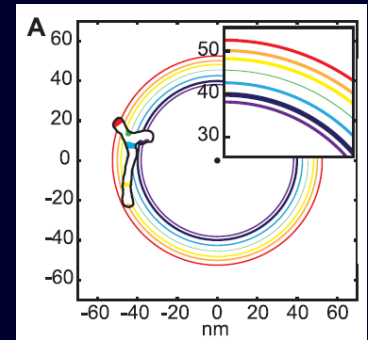


Particle Averaging Improves Resolution of Stereotypic Structures

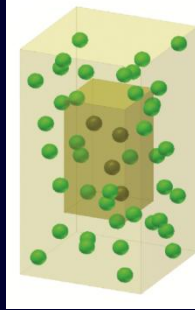
nuclear pore complex proteins



positions
determined
to < 1 nm



Caveats with Super-Resolution Microscopy: Live Cells

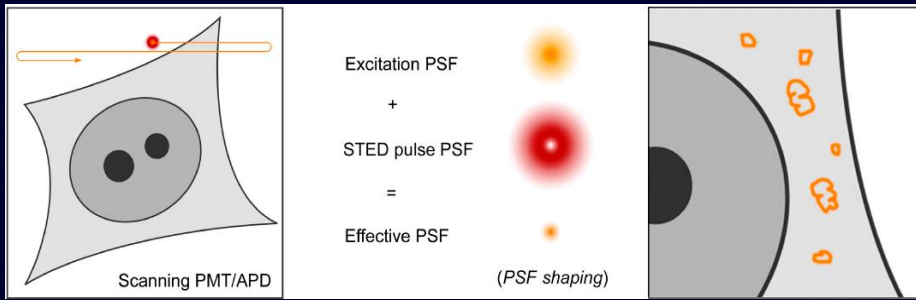


Nyquist criterion:

N -fold resolution increase in D dimensions $\rightarrow N^D$ -fold more photons collected

L. Schermelleh, R. Heintzmann, *J. Cell Biol.* (2010)

STED / RESOLFT



reported resolution (nm)

xy: 20 nm

xyz: 30 nm

photon increase required

100

1,070

intensity (W/cm²)

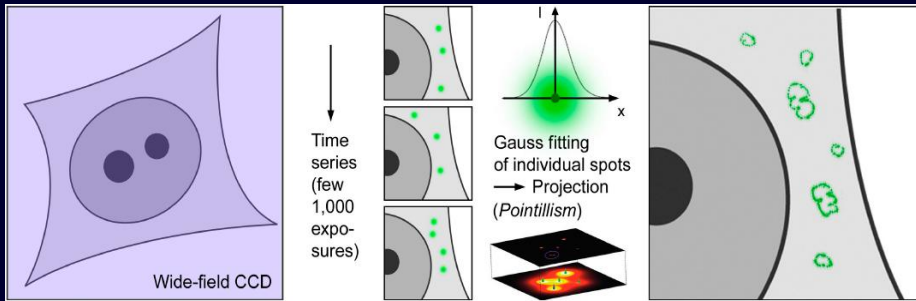
$10^4 - 10^9$

acquisition time (sec)

> 60

~1,000

Localization



xy: 20 nm

xy: 10 nm,
z: 20 nm

100

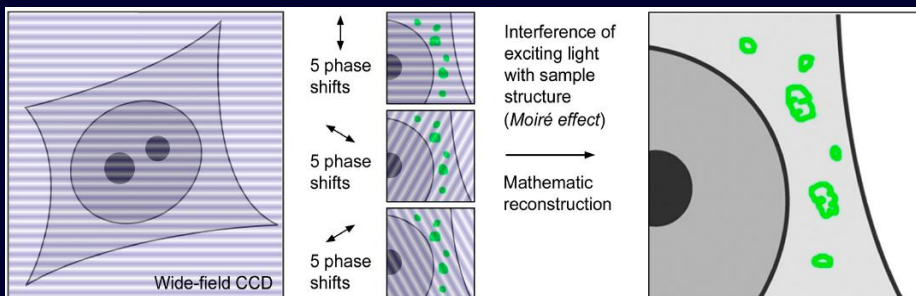
14,400

$10^3 - 10^4$

>20

1,500

SIM



xy: 100 nm

xy: 100 nm,
z: 370 nm

4

8

$10 - 10^2$

0.1 - 1

~10

Live Cell Structured Microscopy

endoplasmic reticulum

2D SIM, 98 nm resolution

0.1 sec acquisition, 1800 frames

Time = 0.00 sec
Frame = 0

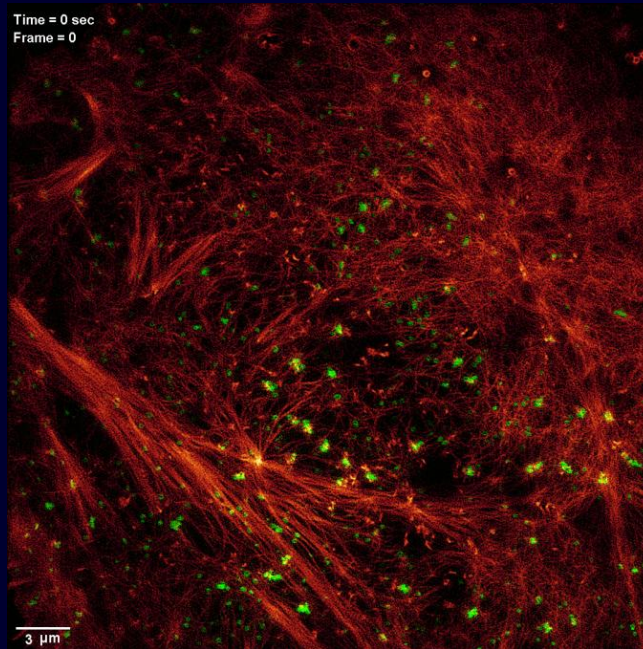


clathrin coated pits and cortical actin

TIRF-SIM, 82 nm resolution

0.5 sec acquisition, 90 frames

Time = 0 sec
Frame = 0

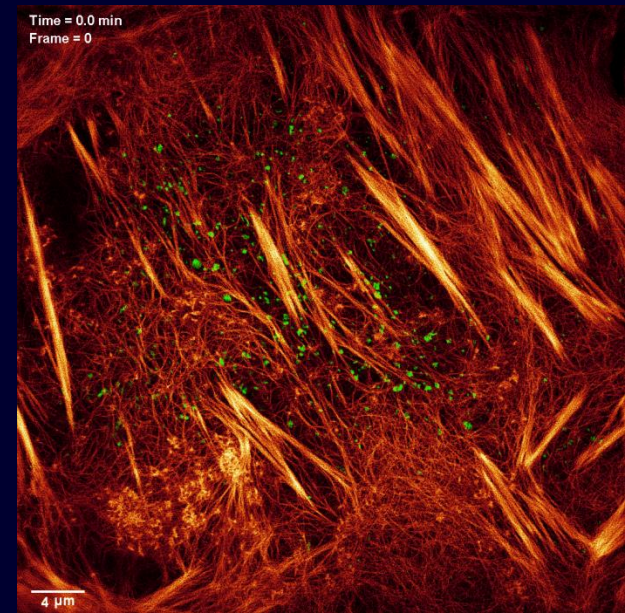


early endosomes and cortical actin

Nonlinear SIM, 62 nm resolution

1.5 sec acquisition, 34 frames

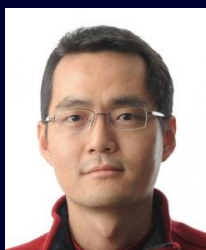
Time = 0.0 min
Frame = 0



Mats
Gustafsson,
1960-2011



Dong
Li



Lin
Shao

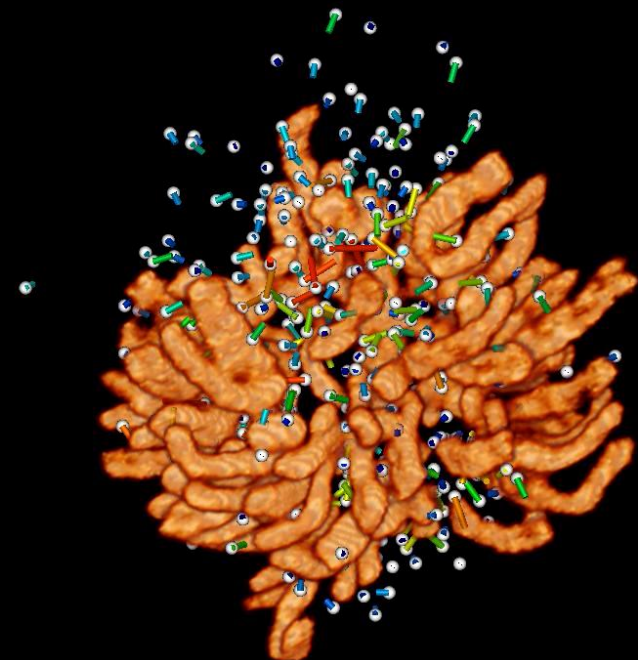
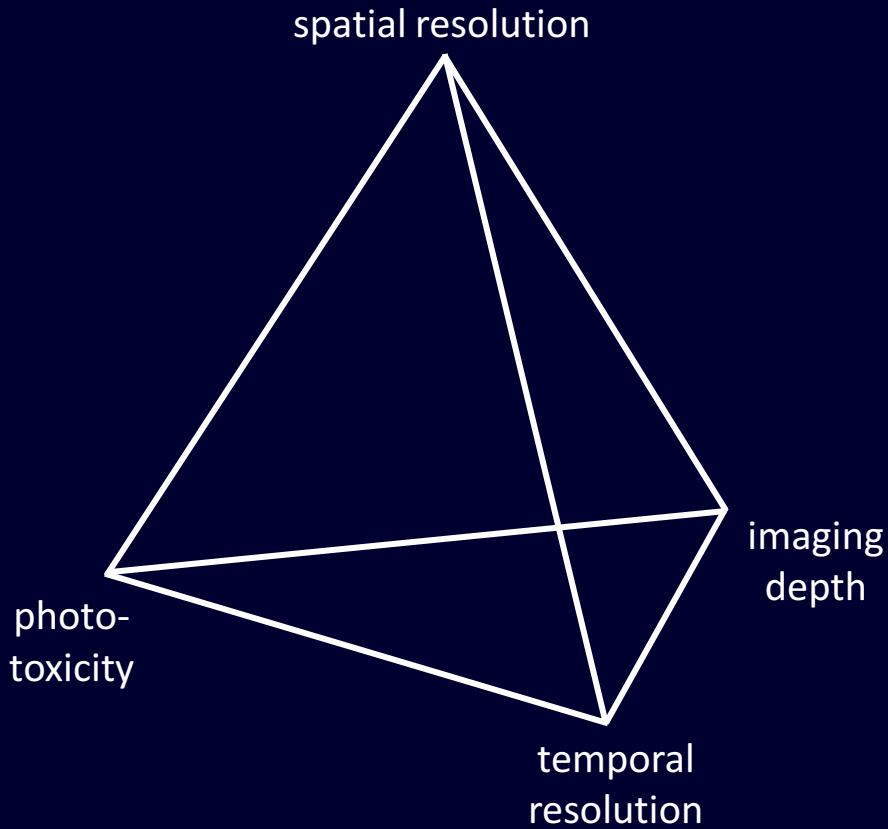
The Challenges and Importance of Studying Live Cell Dynamics

tradeoffs, tradeoffs, tradeoffs

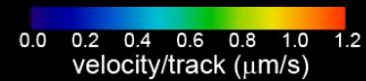
Life is Animate

dividing HeLa cell

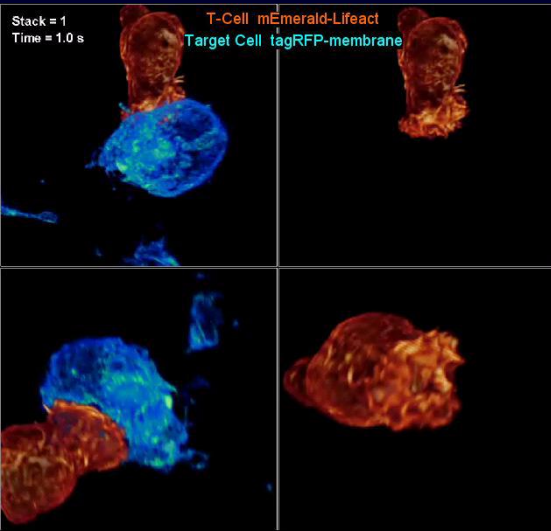
prometaphase



00:00:00

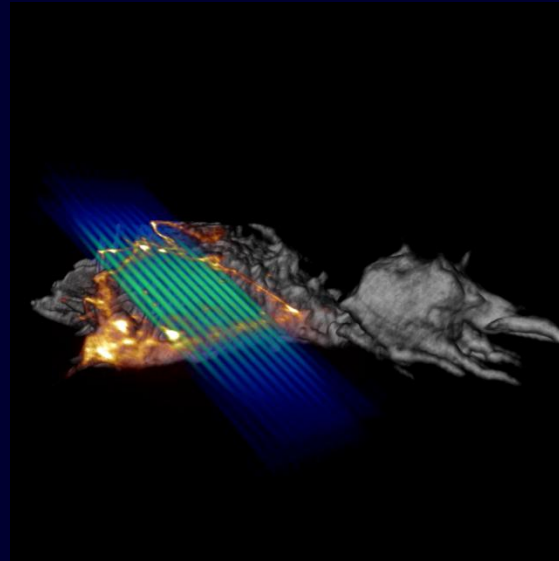


Lattice Light Sheet Microscopy: Non-Invasive 4D Live Cell Imaging

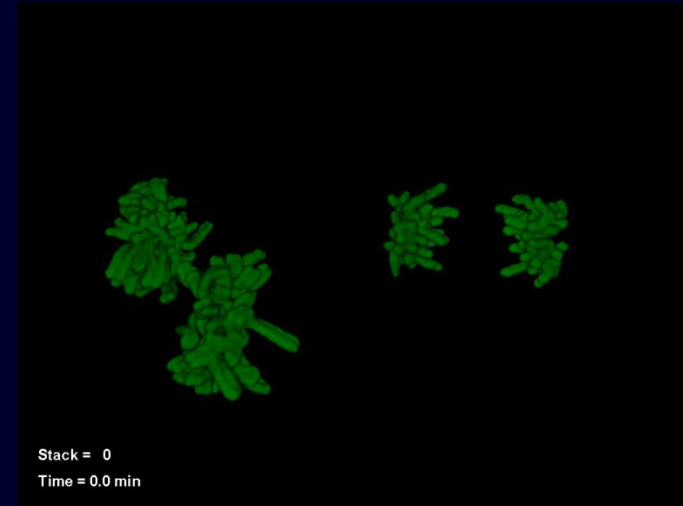


T cell and its target cell

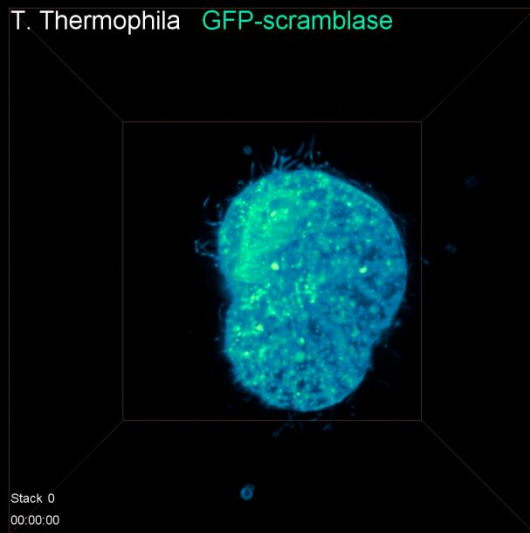
concept



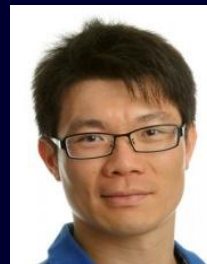
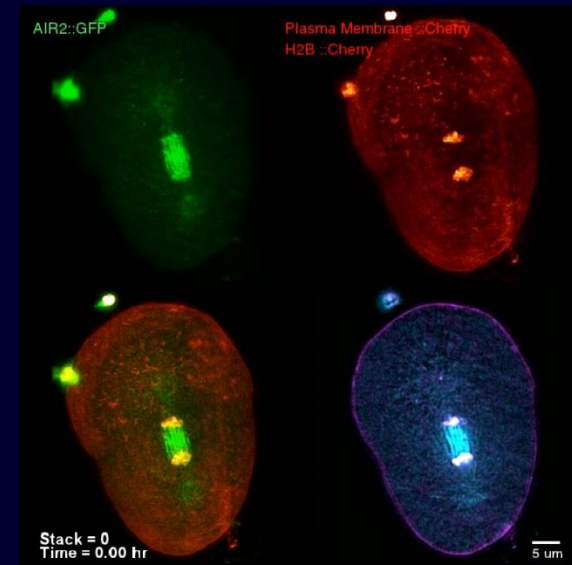
chromosomes, mitosis, and ER during mitosis



Tetrahymena thermophila



C. elegans early embryo



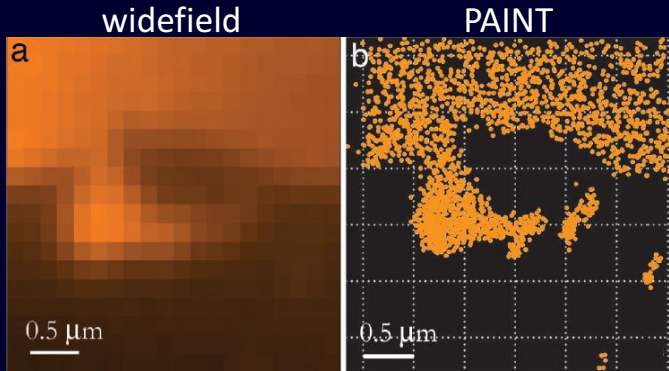
B-C Chen, *et al.*, *Science* **346**,1257998 (2014)

Ultra-High Density 3D Localization Microscopy



Wesley Legant

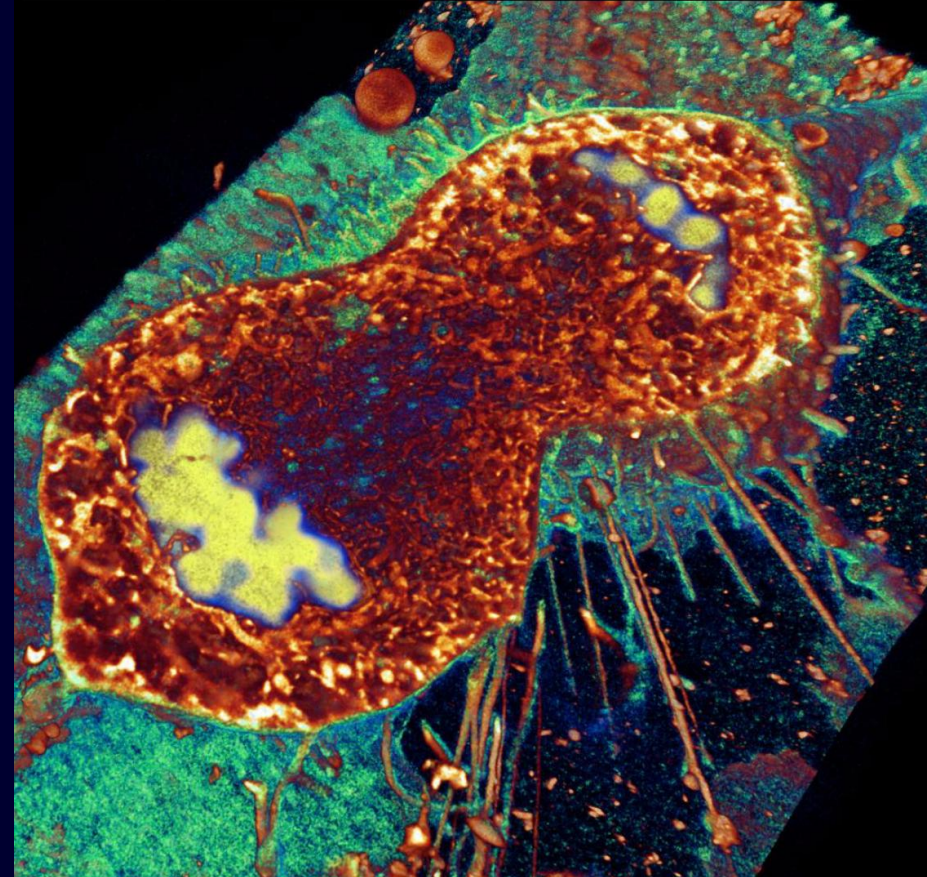
Points Accumulation for Imaging in Nanoscale Topography (PAINT)



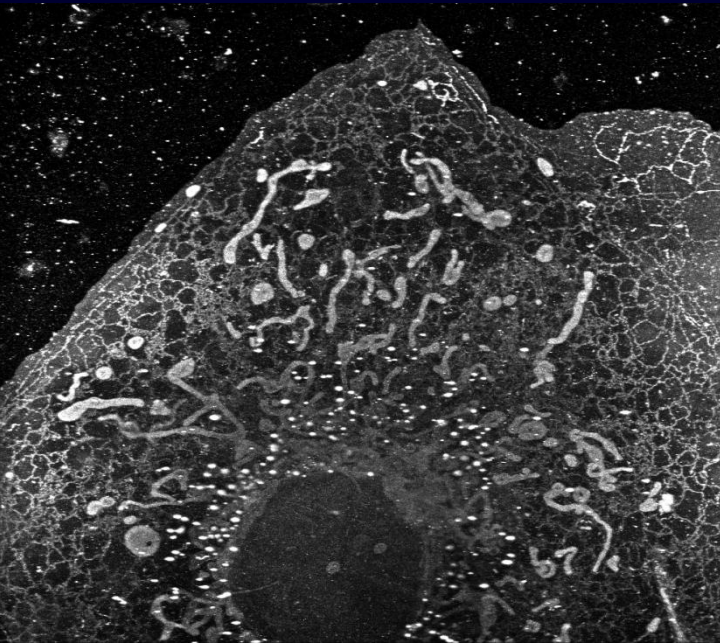
A. Sharonov, R.M. Hochstrasser, *PNAS* 103, 18911 (2006)

3D PAINT with lattice: dividing cell

LLCPK1 Cell Intracellular Membranes Plasma Membrane Histone H2B



over 300 million localized molecules



intracellular membranes, COS-7 cell

Adaptive Optics (AO): Moving Cell Biology Away from the Cover Slip

non-scattering
media: zebrafish
embryonic brain

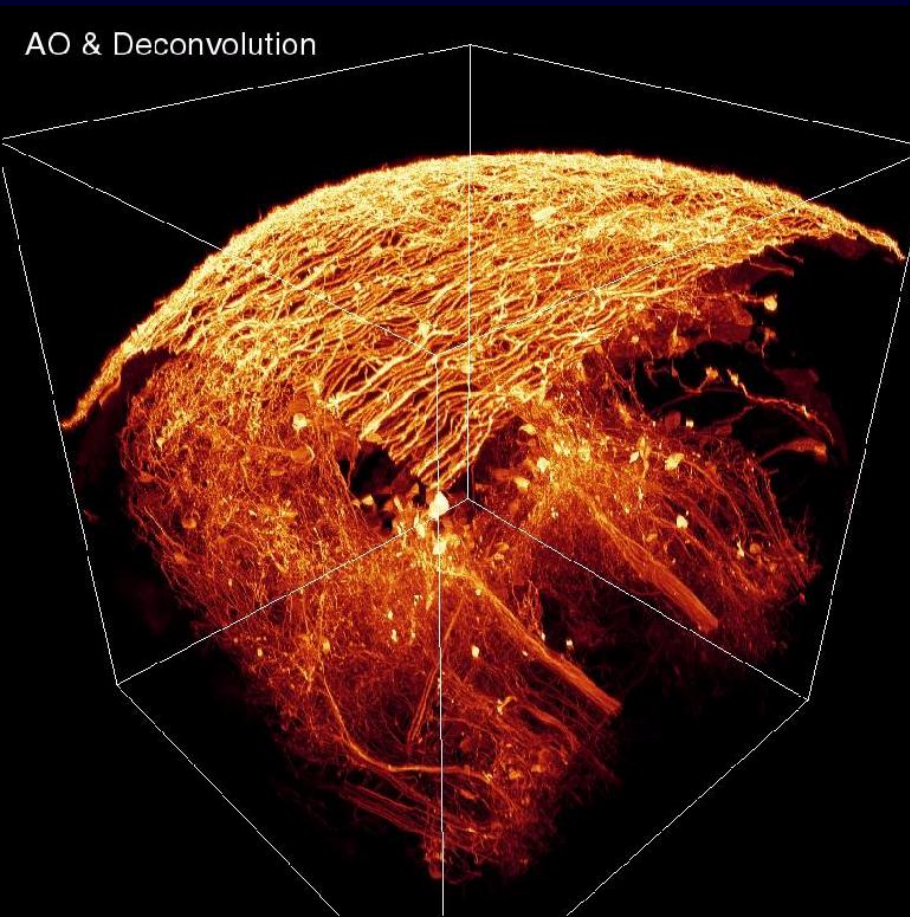


Kai Wang

scattering media:
mouse visual
cortex



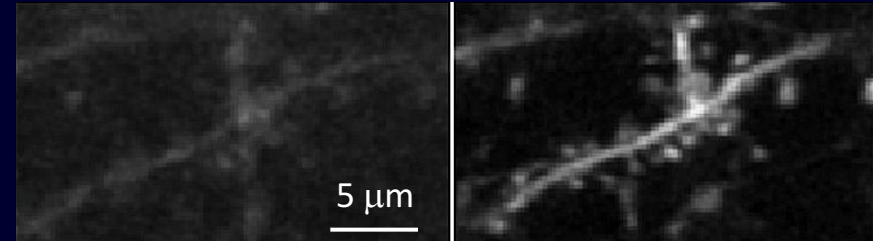
Na Ji



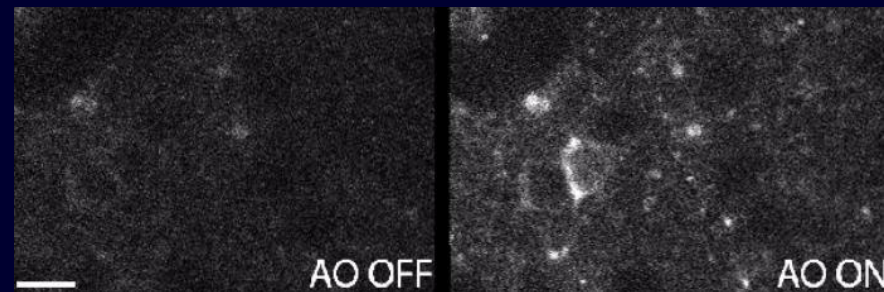
dendritic spines, 600 μm deep

AO off

AO on



functional imaging of neural activity,
400 μm deep



The Beauty and Complexity of Living Systems

